Screening Assessment Report

Ethane, 1,2-dibromo-(1,2-Dibromoethane)

Chemical Abstracts Service Registry Number 106-93-4

Environment Canada Health Canada

Juin 2013

Synopsis

Pursuant to section 74 of the *Canadian Environmental Protection Act, 1999* (CEPA 1999), the Ministers of the Environment and of Health have conducted a screening assessment of Ethane, 1,2-dibromo- (1,2-dibromoethane), Chemical Abstracts Service Registry Number (CAS¹RN) 106-93-4.1,2-Dibromoethane was identified as a priority for assessment because it met the criteria for persistence and/or bioaccumulation and inherent toxicity to non-human organisms. It was also identified as a priority on the basis of greatest potential for human exposure.

1,2-Dibromoethane is considered to be predominantly anthropogenic in origin, though detection of 1,2-dibromoethane in marine air and water suggests possible natural formation as the result of macroalgae growth. In Canada, 1,2-dibromoethane is solely used as a lead scavenger in leaded gasoline for high-performance competition vehicles and piston engine aircraft. Internationally, 1,2-dibromoethane may be used as a grain fumigant; moth control agent in beehives; wood preservative in the timber industry; activator of magnesium in the preparation of Grignard reagents; chemical intermediate in the production of vinyl bromide, plastic and latex; and in the formulation of flame retardants, polyester dyes, resins and waxes. Based on a survey issued under section 71 of CEPA 1999, between 10 000 and 100 000 kg of 1,2-dibromoethane were imported into Canada in the 2000 calendar year.

According to the available information, 1,2-dibromoethane does not degrade quickly in air, and it has a high potential for long-range transport in this medium. It also does not degrade quickly in groundwater. Low experimental bioconcentration factor values suggest that 1,2-dibromoethane has limited bioaccumulation potential in organisms. Therefore, 1,2-dibromoethane meets the criteria for persistence but not for bioaccumulation potential as set out in the *Persistence and Bioaccumulation Regulations*. In addition, experimental toxicity data for 1, 2-dibromoethane suggests that this substance is not expected to cause acute harm to aquatic organisms at low concentrations.

In Canada, 1,2-dibromoethane is routinely monitored in ambient air but not in water, soil or sediments. Risk characterization using conservative exposure concentrations measured in ground water and soil from industrial and non-industrial sites, as well as modelled concentrations for surface water, and critical toxicity values for aquatic and soil organisms indicates that 1,2-dibromoethane is unlikely to cause ecological harm.

Based on the information available with regard to the environment, it is concluded that 1,2-dibromoethane is not entering the environment in a quantity or concentration or under conditions that have or may have an immediate or long-term harmful effect on the environment or its biological diversity or that constitute or may constitute a danger to the environment on which life depends.

¹The Chemical Abstracts Service (CAS) Registry Number is the property of the American Chemical Society, and any use or redistribution, except as required in supporting regulatory requirements and/or for reports to the government when the information and the reports are required by law or administrative policy, is not permitted without the prior written permission of the American Chemical Society.

A critical effect for the characterization of risk of 1,2-dibromoethane exposure to human health is carcinogenicity, as there is strong evidence of carcinogenicity of 1,2-dibromoethane in rats and mice following oral or inhalation exposure. 1,2-Dibromoethane was also genotoxic in several *in vivo* and *in vitro* assays. Therefore, although the mode of induction of tumours has not been fully elucidated, it cannot be precluded that the tumours observed in experimental animals resulted from direct interaction of 1,2-dibromoethane with genetic material.

As mentioned, the sole use of 1,2-dibromoethane in Canada is as a lead scavenger in leaded gasoline for specialized applications. Increases in the releases of this substance to the environment from leaded gasoline are not anticipated, as recent data suggests that the use quantities of these fuels are not increasing. Extensive outdoor and indoor air monitoring data exists for this substance. Although, the substance has occasionally been detected at very low levels, it was not detected in > 99% of the samples analyzed from recent studies. No consumer products containing 1,2-dibromoethane were identified in Canada, and thus exposure from use of consumer products is not expected.

On the basis of the use pattern of 1,2-dibromoethane and the very limited potential for general population exposure, it is concluded that 1,2-dibromoethane is not entering the environment in a quantity or concentration or under conditions that constitute or may constitute a danger in Canada to human life or health.

Based on available information for environmental and human health considerations, it is concluded that 1,2-dibromoethane does not meet one or more of the criteria set out in section 64 of CEPA 1999.

Because this substance is listed on the Domestic Substances List, it is not subject to notification under the *New Substance Notification Regulations (Chemicals and Polymers)*. However, given its hazardous properties, there is concern that new activities that have not been identified or assessed under CEPA 1999 could lead to this substance meeting the criteria set out in section 64 of the Act. Therefore, it is recommended to amend the Domestic Substances List, under subsection 87(3) of the Act, to indicate that subsection 81(3) of the Act applies with respect to this substance, so that any significant new activity is notified and undergoes ecological and human health risk assessments before the substance is imported, manufactured or used for the significant new activity.

Introduction

This screening assessment was conducted pursuant to section 74 of the *Canadian Environmental Protection Act, 1999* (CEPA 1999) (Canada 1999). This section of the Act requires that the Ministers of the Environment and of Health conduct screening assessments of substances that satisfy the categorization criteria set out in section 73 of the Act in order to determine whether they meet or may meet the criteria set out in section 64 of the Act.

Screening assessments focus on information critical to determining whether a substance presents, or may present, a risk to the environment or to human health, according to the criteria set out in section 64 of CEPA 1999. Screening assessments examine scientific information and develop conclusions by incorporating a weight-of-evidence approach and precaution.²Ethane, 1,2-dibromo- (1,2-dibromoethane), CAS RN (Chemical Abstracts Service Registry Number) 106-93-4 was identified as a priority for assessment because it met the criteria for persistence and/or bioaccumulation and inherent toxicity to non-human organisms and was also identified on the basis of greatest potential for human exposure.

The 2004 version of the *State of the Science Report for a Screening Health Assessment* of 1,2-dibromoethane was posted on the Health Canada website on November 29, 2004(Health Canada 2004). The *State of the Science Report for a Screening Health Assessment* was externally reviewed by staff of Toxicology Advice and Consulting Limited and by V.C. Armstrong (consultant) for adequacy of data coverage and defensibility of the conclusions. The external comments were taken into consideration in drafting the *State of the Science Report*. The health screening assessment included here is an update of the *Science Report*.

This screening assessment includes consideration of information on chemical properties, hazards, uses and exposure. Data relevant to the screening assessment of this substance were identified in original literature, review and assessment documents and stakeholder research reports and from recent literature searches, up to January 2010 for ecological sections of the document and September 2009 for human health sections of the document. Also, Canadian monitoring studies, initially reported from draft reports, were updated in this assessment based on finalized reports published in 2010, and another Canadian monitoring study published in 2012, has been included. In addition, an industry survey was conducted in

²A determination of whether one or more of the criteria of section 64 are met is based upon an assessment of potential risks to the environment and/or to human health associated with exposures in the general environment. For humans, this includes, but is not limited to, exposures from ambient and indoor air, drinking water, foodstuffs, and the use of consumer products. A conclusion under CEPA 1999 is not relevant to, nor does it preclude, an assessment against the hazard criteria specified in the *Controlled Products Regulations*, which is part of the regulatory framework for the Workplace Hazardous Materials Information System [WHMIS] for products intended for workplace use. Similarly, a conclusion based on the criteria contained in section 64 of CEPA 1999 does not preclude actions being taken under other sections of CEPA 1999 or other Acts.

2001 through a *Canada Gazette* notice issued under authority of section 71 of CEPA 1999 (Canada 2001). This survey collected data on the Canadian manufacture and import of substances selected for the DSL screening assessment pilot project (Environment Canada 2001a). Key studies were critically evaluated; modelling results may have been used to reach conclusions. When available and relevant, information presented in hazard assessments from other jurisdictions was considered. This screening assessment does not represent an exhaustive or critical review of all available data. Rather, it presents the most critical studies and lines of evidence pertinent to the conclusion.

Evaluation of risk to human health involves consideration of data relevant to estimation of exposure (non-occupational) of the general population, as well as information on health hazards (based principally on the weight-of-evidence assessments of other agencies that were used for prioritization of the substance). Decisions for human health are based on the nature of the critical effect and/or margins between conservative effect levels and estimates of exposure, taking into account confidence in the completeness of the identified databases on both exposure and effects, within a screening context.

This final screening assessment was prepared by officials in the Existing Substances Programs at Health Canada and Environment Canada. As mentioned above, the *State of the Science Report* for a screening health assessment was also previously externally reviewed. The ecological component of this assessment has undergone external written scientific peer review/consultation and comments received were considered in the production of this report. Comments on the technical portions relevant to human health were received from Ms. Joan Strawson, Toxicology Excellence for Risk Assessment, Dr. Michael Jayjock, The LifeLine Group, and Dr. Susan Griffin, U.S. Environmental Protection Agency (EPA). Additionally, the draft of this screening assessment was published on December 16, 2011, subject to a 60-day public comment period and to commenting via the OECD Cooperative Chemicals Assessment Programme. Although external comments were taken into consideration, the final content and outcome of the screening assessment remain the responsibility of Health Canada and Environment Canada.

The critical information and considerations upon which this assessment is based are summarized below.

Substance Identity

Information on the identity of 1,2-dibromoethane is presented in Table 1.

CAS RN	106-93-4				
DSL name	Ethane, 1,2-dibromo-				
	1,2-Dibromoethane (DSL, ECL, EINECS)				
National Chemical	Ethane, 1,2-dibromo- (AICS, ASIA-PAC, DSL, ENCS, NZIoC,				
Inventories names	PICCS, SWISS, TSCA)				
	Ethylene dibromide (PICCS)				
Other names	Aadibroom, Bromofume, α , β -Dibromoethane, α , ω -Dibromoethane,				
Other names	Ethylene dibromide,				
Chemical group (DSL	Discrete organics				
stream)					
Major chemical class	Alkanes				
or use					
Major chemical	Halogenated alkanes				
subclass					
Chemical formula	$C_2H_4Br_2$				
Chemical structure	H H Br—C—C—Br H H				
SMILES	C(CBr)Br				
Molecular mass	187.86 g/mol				

Abbreviations: AICS, Australian Inventory of Chemical Substances; ASIA-PAC, Asia-Pacific Substances Lists; CAS RN, Chemical Abstracts Service Registry Number; DSL, Domestic Substances List; ECL, Korean Existing Chemicals List; EINECS, European Inventory of Existing Commercial Chemical Substances; ENCS, Japanese Existing and New Chemical Substances; NZIoC, New Zealand Inventory of Chemicals; PICCS, Philippine Inventory of Chemicals and Chemical Substances; SMILES, Simplified Molecular Input Line Entry Specification; SWISS, Swiss Giftliste 1 and Inventory of Notified New Substances; TSCA, *Toxic Substances Control Act* Chemical Substance Inventory. Source: National Chemical Inventories (2006).

Physical and Chemical Properties

Physical and chemical properties identified for 1,2-dibromoethane are summarized in Table 2 below.

Property ¹	Туре	Value ²	Temperature
Melting point (°C)	Experimental	9.9	-
Boiling point (°C)	Experimental	131.6	-
Vapour pressure (Pa)	Experimental	1493	25°C
		(11.2 mmHg)	
Henry's Law constant	Experimental	65.9	25°C
(Pa·m ³ /mol)		$(6.50 \times 10^{-4} \text{ atm} \cdot \text{m}^3/\text{mol})$	
Log K _{ow} (dimensionless)	Experimental	1.96	_
$Log K_{oc}^{3}$ (dimensionless)	Estimated	1.70 (K _{ow} method)	_
Water solubility (mg/L)	Experimental	3910	25°C
k_{OH} (cm ³ /molecule per	Experimental	$2.50 imes 10^{-13}$	25°C
second)			

Abbreviations: K_{oc} , organic carbon-water partition coefficient; k_{OH} , rate constant for gas-phase reaction with hydroxyl radical; K_{ow} , octanol-water partition coefficient.

¹ All physical and chemical properties were obtained from the Syracuse Research Corporation's PhysProp database (PhysProp 2009) except as noted.

² Values in parentheses are original values given in the database.

³ The estimated value was calculated by PCKOCWIN (2008).

Sources

1,2-Dibromoethane is considered to be predominantly anthropogenic in origin, although its detection in marine air and water suggests possible natural formation as the result of macroalgal growth (Class and Ballschmiter 1988). Commercial production involves an exothermic reaction of liquid bromine and gaseous ethene in a glass reactor column packed with coiled heat exchangers (Gerhartz 1985). Synthesis of 1,2-dibromoethane is also possible using acetylene (ethyne) and hydrobromic acid as starting materials (Budavari et al. 2001).

Based on responses to a survey issued under section 71 of CEPA 1999, between 10 000 and 100 000 kg of 1,2-dibromoethane were reported to be imported into Canada in the 2000 calendar year for use as a fuel additive (Environment Canada 2001a). Such quantity indicates a considerable decrease from the 11 million kilograms reported during the period of compilation of the DSL (1984–1986).

1,2-Dibromoethane was also reported to be manufactured in or imported into Canada in the 2000 calendar year, in a mixture of a product at a low concentration (< 1% w/w); however the total quantity of 1,2-dibromoethane in the product at a low concentration (<1% w/w) in 2000 was unknown.

Uses

Based on responses to a survey issued under section 71 of CEPA 1999, 1,2dibromoethane is solely used in Canada as a lead scavenger to prevent build-up of lead oxide in engines running on leaded gasoline (Environment Canada 2001a). Leaded gasoline was banned in cars in 1990 when the *Gasoline Regulations* came into force under CEPA (Canada 1990) and was further phased out after an amendment to the Regulations removing an exemption on leaded gasoline used in farm machinery, boats and trucks over 3856 kg in April 2008. This reduction in use of leaded gasoline coincides with the reduction in import volumes of 1,2-dibromoethane in Canada from the period of compilation of the DSL (1984–1986) to the 2000 calendar year. Presently, 99.8% of gasoline used in Canada is unleaded (Environment Canada 2009a).

The *Gasoline Regulations* do not apply to leaded gasoline for aviation. In addition, the Regulations allow for leaded gasoline use in competition vehicles (Canada 2010). Use of leaded gasoline in aircraft represented 98% of total leaded fuel in Canada in 2009, while high-performance competition vehicles represented 2% (June 2009 email from Oil, Gas and Alternative Energy Division, Environment Canada, to Existing Substances Risk Assessment Bureau, Health Canada; unreferenced). Leaded aviation gasoline represents a small percentage (approximately 1.5%) of total aircraft fuel in Ontario (Patriarche and Campbell 1999).

1,2-Dibromoethane was introduced worldwide as a soil and grain fumigant in 1946. Canada and the United States discontinued its use in pesticide products in 1984, and it was subsequently banned as an agricultural pesticide in member states of the European Union and many other countries (Packer 1980; UNEP and FAO 2003; PPDB 2009). There is evidence that 1,2-dibromoethane may still be applied by some individual beekeepers in Greece to control moth infestations in honeycombs (Tananaki et al. 2005, 2006). In addition, 1,2-dibromoethane may be used as a wood preservative against pest damage in Australia, and therefore post-application residues of 1,2-dibromoethane in imported wood and wood products may exist (NPI 2006).Today, 1,2-dibromoethane is listed under the Prior Informed Consent (PIC) procedure of the Rotterdam Convention, 1998, under the sponsorship of the United Nations Food and Agriculture Organization and the United Nations Environment Programme (UNEP and FAO 2003).

Globally, 1,2-dibromoethane is used principally as a chemical intermediate and industrial solvent. Uses include activation of magnesium in the preparation of Grignard reagents; use as a chemical intermediate in the production of plastic, latex, and vinyl bromide; a flame retardant used in modacrylic fibres; and use in the formulation of polyester dyes, resins and waxes (HSDB 2010; NTP 2005). As no chemical reaction is completely efficient, some 1,2-dibromoethane may remain as an unintended manufacturing residue in articles.

Use of 1,2-dibromoethane in consumer products has not been identified.

1,2-Dibromoethane is not expected to be present in cosmetic products in Canada, as it is not listed as an ingredient in the Cosmetic Notification System database (CNS 2009). There are no registered pesticides that contain 1,2-dibromoethane as an active ingredient or formulant in Canada (PMRA 2007), and 1,2-dibromoethane is not listed as an approved food additive under Division 16 of the *Food and Drug Regulations* (Canada [1978]).

Releases to the Environment

1,2-Dibromoethane is not reportable to Canada's National Pollutant Release Inventory (NPRI 2008). According to the United States Toxics Release Inventory Program, total on-site and off-site disposal or other releases of 1,2-dibromoethane in the 2007 calendar year amounted to 1921 kg, where 1686 kg were released as fugitive air emissions, 96 kg as point source air emissions, 0.45 kg as surface water discharges and 0 kg as land treatment (TRI 2007). These release details suggest that air may also be the primary receiving compartment of 1,2-dibromoethane releases in Canada.

1,2-Dibromoethane will primarily enter the atmosphere from fugitive emissions associated with its use as a scavenger in leaded gasoline, which converts lead oxides to lead halides (ATSDR 1992). Some of the 1,2-dibromoethane is broken down during the scavenging process and some is emitted in the non-transformed form (IPCS 1996). Methyl bromide is also emitted. According to the US EPA (1999), the 1,2-dibromoethane emissions from mobile sources were estimated to be equal to zero. Therefore, engine exhaust releases of the substance are likely negligible and most of the releases come from fugitive emissions like spills, leaks and evaporation from reservoirs containing leaded gasoline. Evaporative losses can also occur during refilling and transfers. Based on the 1999 Inventory of Toxic Air Emissions for the Great Lakes states and the province of Ontario (Great Lakes Commission 2002), releases of 1,2-dibromoethane were estimated at 10.69 pounds/year (4.86 kg) from point sources (separately identified device/process at each facility source) and at 13.34 pounds/year (6.06 kg) for area sources (aggregation of similar or identical devices/processes within a defined area) for a total of 24.03 pounds (10.92 kg) released in 1999. No other anthropogenic release information in Canada has been found for 1,2-dibromoethane.

In addition, 1,2-dibromoethane appears to be formed naturally by microalgae growth and has been detected in ocean waters and air (IRIS 2002). Laturnus (1996) mentions that Arctic brown, red and green macroalgae release volatile halogenated organic compounds including 1,2-dibromoethane. The extent of the contribution of these natural sources to global emissions is unknown. Class and Ballschmiter (1988) found baseline concentrations of 1,2-dibromoethane in air (20 ng/m³) and in marine waters (0.02 ng/L) collected from open areas of the North and South Atlantic Ocean. The source of the compound could be the natural production by algae and/or the anthropogenic emissions.

Environmental Fate

Environmental fate analysis integrates information on the chemical behaviour of the substance with the properties of the receiving environment. The objective of fate analysis is to determine the multimedia distribution of the substance after its release into the environment. This includes consideration of the persistence and bioaccumulation of the substance in the environment.

Based on its physical and chemical properties (Table 2), the results of Level III fugacity modelling (Table 3) suggest that 1,2-dibromoethane is expected to predominantly reside in air, water or soil depending on the compartment of release.

	Percentage of substance partitioning into each compartment				
Substance released to:	Air	Water	Soil	Sediment	
Air (100%)	93.7	5.28	0.90	0.072	
Water (100%)	13.7	85.9	0.141	0.327	
Soil (100%)	14.4	6.54	79	0.025	

Table 3. Results of the Level III fugacity modelling (EQC 2003)

If released to air, a high amount of the substance is expected to reside in air (see Table 3 above). Based on the high vapour pressure of 1493 Pa and the moderate Henry's Law constant of $65.86 \text{ Pa} \cdot \text{m}^3/\text{mol}$, 1,2-dibromoethane is volatile. Therefore, if released solely to air, it will tend to reside in this compartment (~94%, see Table 3).

If released into water, 1,2-dibromoethane is expected to weakly adsorb to suspended solids and sediment based upon its low log K_{oc} value of ~1.70. Volatilization from water surfaces is expected to be a moderate fate process based upon this compound's experimental Henry's Law constant. Thus, if water is a receiving medium, 1,2-dibromoethane is expected to mainly reside in water and to some extent partition to air (see Table 3).

If released to soil, 1,2-dibromoethane is expected to have moderate adsorptivity to soil and is expected to be fairly mobile based upon its estimated log $K_{oc.}$ Volatilization from moist soil surfaces seems to be a moderate fate process based upon its experimental Henry's Law constant. This chemical may volatilize from dry soil surfaces based upon its high vapour pressure. Therefore, if released to soil, 1,2-dibromoethane will mostly reside in this environmental compartment, and also partition to water and air, as illustrated by the results of the Level III fugacity modelling (see Table 3).

These results represent the partitioning of the substance in a hypothetical evaluative environment resulting from intermedia partitioning, and loss by both advective transport (out of the modelled region) and degradation/transformation processes. The partitioning values shown in Table 3 represent the net effect of these processes under conditions of continuous release when a non-equilibrium "steady-state" has been achieved. In addition, the Transport and Persistence Level III Model (TaPL3) version 3 (TaPL3 2000) has been used to estimate a characteristic travel distance (CTD) for 1,2dibromoethane in air, defined as the maximum distance traveled in air by 63% of the substance. The CTD of the substance was 51 022 km. Furthermore, Beyer et al. (2000) have defined 3 classes estimating the potential for long-range transport in air according to the CTD: class 1 (long CTD) > 2000 km; class 2 (intermediate CTD) 700-2000 km and class 3 (short CTD) < 700 km. Therefore, 1,2-dibromoethane belongs to the class 1 and is considered to have a high potential for long-range transport in air.

Persistence and Bioaccumulation Potential

Environmental Persistence

1,2-Dibromoethane degrades very slowly in the atmosphere. It degrades by reaction with photochemically produced hydroxyl radicals, with a half-life of 64 to 69 days (IUCLID 2000). A half-life of 138 days was calculated by Qiu et al. (1992) in the troposphere. According to Howard et al. (1991), the photo-oxidation half-life in air is between 10.7 and 107 days.

1,2-Dibromoethane has been found to degrade both aerobically and anaerobically (ATSDR 1992; Falta et al. 2005). In surface waters, the main removal process for 1,2-dibromoethane is by volatilization, with a half-life within 1 to 5 days (IUCLID 2000) and up to 16 days (ATSDR 1992). 1,2-Dibromoethane is not expected to readily volatilize from water; however, it can evaporate from the free-phase gasoline (Falta et al. 2005). Little degradation of 1,2-dibromoethane occurs through direct photolysis in water, as evidenced by the half-life greater than 1 year (IUCLID 2000).

In groundwater, where volatilization is not possible, studies have shown that 1,2-dibromoethane can persist for years (Pignatello and Cohen 1990). In Florida groundwater, the substance has been determined to have a chemical half-life of 1.5 to 2 years at 22°C (Weintraub et al. 1986). In addition, 1,2-dibromoethane tends to be mobile in groundwater due to its low octanol-water partition coefficient (Falta et al. 2005). Hydrolysis is the major mode of degradation, giving ethylene glycol and bromide ion (Weintraub et al. 1986). In the laboratory, Vogel and Reinhard (1986) estimated a hydrolysis half-life for 1,2-dibromoethane of 2.5 years in water at 25°C and pH 7. Half-lives as long as 354 days to 13.2 years are reported for the water compartment in IUCLID (2000). Finally, Howard et al. (1991) reported a half-life interval of 28 to 180 days for surface waters and of 19.6 to 120 days for groundwater. These values are lower than the other sources possibly because of the influence of biotic degradation in addition to abiotic degradation. As a result of its hydrolytic stability and the limited biological activity in subsurface soils, 1,2-dibromoethane leached to groundwater is expected to persist for years (ATSDR 1992). Uncertainty about the mechanism and rates of biotic and abiotic degradation poses a challenge to the understanding of the subsurface fate and transport of this substance (Falta et al. 2005).

In soils, 1,2-dibromoethane is used as a fumigant, and most of the substance is expected to be rapidly lost by volatilization to the atmosphere and leaching to surface waters and groundwater (IPCS 1996). According to results from a study on biodegradation of 1,2-dibromoethane by soil microorganisms, the substance was almost completed degraded within 1 week; however, a small fraction may persist in topsoil for up to several years (Pignatello 1986). It may be due to that the substance could react with nucleophilic O or S groups on soil organic matter, developing any covalent attachment. According to Walton et al. (1989), the degradation half-lives are 3.1 and 1.9 days, in silt loam and sandy loam, respectively.

Partitioning to sediment is not expected to be an important process in the environment because of the low sorption potential, high vapour pressure, and high water solubility for 1,2-dibromoethane. This was shown in the results of the Level III fugacity modelling where the proportion of the substance in sediments at equilibrium is very low (0.1%). For these reasons, persistence in sediments has not been assessed.

Based on the empirical data available and calculated values, 1,2-dibromoethane meets the persistence criteria in air (half-life in air \geq 2 days) and water (half-lives in water \geq 182 days) but does not meet the criteria for soil (half-life in soil \geq 182 days) as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

Potential for Bioaccumulation

An experimental log K_{ow} value of 1.96 for 1,2-dibromoethane suggests that this chemical has low potential to bioaccumulate in biota (see Table 2).

Bioconcentration factor (BCF) data exist in the literature, but no bioaccumulation factor (BAF) data have been found for 1,2-dibromoethane. BCF values for this substance range from < 1 to 20. Two key studies are reported herein.

A mean BCF value of 2.7 was found for a nematode, *Aphelenchus avenae* (Marks et al. 1968). Individuals were exposed for 30 minutes at 4°C and 20°C to the following three concentrations of 1,2-dibromoethane: 488, 996 and 1991 mg/L. The authors also reported BCF values of 6, 9 and 20 for other nematode species: *Pellodera* sp., *Tylenchulus semipenetrans* and *Anguina tritici*, respectively. Carp (*Cyprinus carpio*) exposed to an aqueous solution containing 1,2-dibromoethane at 15 and 150 μ g/L had a BCF ranging from < 3.5 to 14.9 and 1.6 to 3.2, respectively (CITI 1992).These low BCF values indicate that 1,2-dibromoethane does not bioconcentrate to a great extent in organisms. Therefore, the compound is not expected to bioaccumulate in organisms or biomagnify in food chains.

Based on the empirical data available, 1,2-dibromoethane does not meet the bioaccumulation criterion (BCF and $BAF \ge 5000$) as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

Potential to Cause Ecological Harm

Ecological Exposure Assessment

1,2-Dibromoethane has been detected in ambient air, soils, groundwater and food (ATSDR 1992). Environmental concentrations in the United States have been reported frequently, but Canadian data are scarce. Where available, Canadian ambient environmental concentrations were used in the determination of predicted environmental concentrations (PECs) for the purpose of characterizing ecological risk. Where recent Canadian data were unavailable, predictive models have been used. It should be noted that many concentrations reported in this section were measured in the 1970s and 1980s, when 1,2-dibromoethane was widely used. As it is no longer widely used, this has been taken into account when selecting environmental concentrations to be used as PECs. The details of measured and predicted environmental concentrations of 1,2-dibromoethane are summarized in Appendices 2 to 6.

In Canada, 1,2-dibromoethane is routinely monitored in air, but not in water, soil or sediments. Air measurements in Canada for the years 2004–2009 indicated a maximum concentration of 60 ng/m³ (Environment Canada 2009b), a decrease from the previously measured maximum level of 143 ng/m³ in 2002 (Environment Canada 2004). Additionally, a conservative air concentration of 1,2-dibromoethane was estimated with the SCREEN3 model (SCREEN3 1995); the resulting value was 377.4 ng/m³ (see Appendix 2). This more conservative modelled value was selected as the PEC for air.

For surface waters, the predictive model ChemSim (2003) was run. ChemSim is a geographic information system-based aquatic exposure estimation model designed to estimate the dispersion and transport of substances released to watercourses. ChemSim combines estimated release quantities with information regarding the receiving watercourses to estimate aquatic exposure values (see Appendix 4). The following assumptions were used:

- The effluent release type was continuous from a steady point source.
- 1% of the total amount reported annually by a company was assumed to be the release quantity at one plant or divided among eight facilities. Of this, 10% was assumed to be released surface water either directly or via a sewage treatment plant.
- 10, 25 and 50 percentile flow estimates were considered for the receiving river.
- Sewage treatment plant (STP) removal rates were used for some of the simulations.

The only reported measurement for groundwater in Canada is 5.0 μ g/L (Environment Canada 2001b).

For soils, the highest Canadian concentration was found at a depth of 3 m (see Appendix 6). However, this concentration was not used as the PEC since the shallow subsurface of soil is the most representative zone where soil invertebrates live. Thus, the reported soil measurement of 8×10^{-2} mg/kg dry weight measured in surface soil 0.2-0.76 m deep was used (Environment Canada 2001b).

Ecological Effects Assessment

Several studies relating to acute and chronic toxicity of 1,2-dibromoethane to fish, soil and aquatic invertebrates, and microorganisms were identified and critically reviewed. Studies with the most sensitive and reliable results are discussed below. The appropriate median lethal concentration (LC_{50}) values were selected as critical toxicity values (CTVs) for the purposes of ecological risk characterization.

Chronic toxicity of 1,2-dibromoethane was evaluated in the fruit fly larvae, *Drosophila melanogaster* (Chroust et al. 2007). Fruit flies, kept in glass bottles, were exposed to 1,2-dibromethane by inhalation for 48 hours to induce chronic effects. Ambient concentration of the substance in the experimental bottles was monitored by a gas chromatograph every 12 hours; however, these measurements were not provided. Rather, LC_{50} values resulting from exposure were expressed in μ g/L. For this reason, the LC_{50} value for *Drosophila melanogaster* could not be used in the environmental risk assessment since the risk quotient (RQ) could not be calculated for this exposure scenario.

Holcombe et al. (1995) conducted a flow-through 96-hour acute test using larval Japanese medaka (*Oryzias latipes*). Concentrations of 1,2-dibromoethane were measured throughout the course of experiment. The measured 96-hour LC_{50} was 32.1 mg/L.

The carcinogenicity of the substance towards the same species was investigated by Hawkins et al. (1998). Juvenile fish were chronically exposed to three concentrations in a flow-through system for 73 to 97 days. The measured concentrations in water in the low, intermediate and high concentration groups were 0.13mg/L, 6.20mg/L and 18.58 mg/L, respectively. Samples were taken for histological examination at 24, 36 and 58 weeks from the beginning of the tests. 1,2-Dibromoethane was clearly carcinogenic to the Japanese medaka in the intermediate and high concentration groups, causing: (i) hepatocellular adenomas and carcinomas, (ii) cholangiomas, (iii) cholangiocarcinomas and (iv) gall bladder papillary adenomas and adenocarcinomas.

The authors also evaluated the toxic effects during an approximately 90-day exposure period, by looking at mortalities, fecundity, viable embryos, hatch, fry survival, and abnormal embryos (Hawkins et al. 1998). Toxic responses varied and the trends were not associated with the concentration levels of 1,2-dibromoethane in the test solutions. For the overall survival and fecundity (measured by the total number of viable eggs produced during a 23-day collection period), the intermediate concentration group demonstrated a lower survival (46% of mortality) and lower fecundity (0%) than the low concentration group (0.3% of mortality and 59% of fecundity) and high concentration group (1.1% of mortality and 2% fecundity). Fry survival rate was much lower in the low concentration group (43%) than the control groups (91.9% for the static control, and 84.2% for the

flow-through control), while data was not available for the intermediate and high concentration groups. For the gross abnormal embryos, the low concentration group demonstrated a higher incidence (6.8%)than both control groups, however data were not available either for the intermediate and high concentration groups.

Based on the observation from the study, the flow-through control group didn't demonstrate significantly more toxic effect than the static control, and 0.034 mg/L was considered as no-observed-effect-concentration (NOEC). The very next exposure level, 0.133 mg/L used in the low concentration group, was then considered as the lowest-observed-effect-concentration (LOEC). The resulting maximum acceptable toxicant concentration (MATC), i.e., the concentration falling between the highest concentration showing no effect and the next higher concentration showing a toxic effect when compared to the controls, was 0.067 mg/L, calculated as the geometric mean between the NOEC and LOEC values in the study.

Kszos et al. (2003) evaluated the acute toxicity of 1,2-dibromoethane on three species: larval fathead minnow (*Pimephales promelas*), *Daphnia magna* and *Ceriodaphnia dubia* in a static closed system. The concentrations of 1,2-dibromoethane were monitored during the experiments. The 48-hour LC_{50} for *C. dubia* was 3.61 mg/L, and 6.5 mg/L was reported for *D. magna*. The 96-hour LC_{50} for the fathead minnow was 4.30 mg/L.

Reliable data on aquatic algae were not identified.

Reliable acute or chronic toxicity data have not been found either for soil organisms. Therefore, a quantitative structure-activity relationship (QSAR) model was used to estimate a 14-day LC_{50} value of 330 mg/L for the earthworm (ECOSAR 2008).

No studies have been found for benthic organisms. Had they been available, they would not have been relevant for this assessment, given that 1,2-dibromoethane is unlikely to partition to sediments.

No ecological studies were identified for terrestrial wildlife.

Laboratory studies on mammals have been conducted with 1,2-dibromoethane to evaluate the potential for impacts to human health; relevant data from these studies are presented in the Potential to Cause Harm to Human Health section of this screening assessment.

Characterization of Ecological Risk

The approach taken in the ecological risk characterization is to examine various supporting information and develop conclusions based on a weight-of-evidence approach. Particular consideration has been given to risk quotient analyses, persistence, inherent toxicity, environmental realism of the exposure scenario used to derive PECs, and widespread occurrence in the environment. Endpoint organisms have been selected based on analysis of exposure pathways. For each endpoint, a conservative (reasonable worst-case) PEC and a predicted no-effect concentration (PNEC) are determined. The

PNEC is arrived at by selecting the lowest CTV for the relevant organism and dividing it by an application factor appropriate for the data point. A risk quotient (PEC/PNEC) is calculated for each of the endpoint organisms in order to contribute to the characterization of ecological risk in Canada. A summary of data used in the ecological risk characterization of 1,2-dibromoethane is presented in Table 4.

Assessment endpoints were evaluated in a few different exposure scenarios. CTVs were selected for the most sensitive endpoints from pelagic (aquatic) organisms and soil organisms. CTVs were selected as the lowest critically reviewed literature value for each group of organisms. For pelagic organisms, acute and chronic data were used. Since no reliable acute or chronic toxicity data were found for soil organisms, a modelled value was selected for the aquatic compartment. No studies were identified for benthic organisms, and they are not considered further in this assessment. The CTVs for each group of organisms are presented in Table 4 below.

Assessment endpoint	Organism	CTV	Application factor	PNEC	PEC	Risk quotient (PEC/ PNEC)
Pelagic organisms (reproduction)	Japanese medaka (Oryzias latipes)	67 μg/L (chronic) ^a	10	6.7 μg/L	5.0 μg/L (groundwater, see Appendix 4)	0.75
					2-3 μg/L ^b (surface water, see Appendix 4)	0.30-0.45 ^b
Soil organisms (mortality)	Earthworm (modelled value)	329.75 mg/kg dry weight for soil ^c	100	3.3 mg/kg dry weight for soil	8×10^{-2} mg/kg dry weight (industrial site, see Appendix 6)	2.4 × 10 ⁻²
					3.9×10^{-4} mg/kg dry weight (non- industrial site, see Appendix 6)	1.2 × 10 ⁻⁴

 Table 4. Summary of data used in the risk characterization of 1,2-dibromoethane

^a Hawkins et al. 1998.

^b Based on ChemSim (2003) simulations with realistic scenarios: the total amount reported is divided among eight facilities. Concentrations and risk quotients calculated at a distance of 50 m downstream from the point of discharge with 10% flow and with or without sewage treatment plant (STP) removal. At a distance of 10 m from the point of discharge, risk quotients are still below 1.

² CTV for soil organisms was calculated using modelled an LC₅₀ value of 329.75mg/L for earthworm (ECOSAR 2008) with application of the equilibrium partitioning equation (EqP) (Environment Canada 1996) as follows: $CTV_s = CTV_i \times f_{oc} \times K_{oc}$ where:

 CTV_i = critical toxicity value for invertebrates (329.75mg/L)

 f_{oc} = mass fraction of organic carbon in the solid phase (0.02 default value for soil) K_{oc} = organic carbon partition coefficient (10^{1.7} = 50, log K_{oc} =1.70, Table 2, 10^{1.7} = 50 is an average from Table 2)

For pelagic organisms, Japanese medaka (*Oryzias latipes*) was the most sensitive to 1,2-dibromoethane, with a maximum acceptable tolerable concentration of 67 μ g/L for chronic effects (reproduction). This value was chosen as the CTV; it may be most representative and realistic of the threshold where chronic effects might occur. The CTV was divided by a factor of 10 to account for extrapolation from laboratory to field conditions and interspecies and intraspecies variations in sensitivity, resulting in the PNEC of 6.7 μ g/L.

For the characterization of risk to pelagic organisms, PEC values were selected to represent exposure from surface waters and from groundwater. For surface waters, the lowest values predicted by ChemSim simulation (ChemSim 2003) using a point source release scenario were used as PEC. For groundwater, it is assumed that the contaminated groundwater will recharge surface waters; therefore, potential effects are examined using pelagic species. Thus, the measured groundwater concentration of $5.0 \mu g/L$ was used as the PEC for estimating potential risk from the seepage of contaminated groundwater to surface water.

The risk quotient (RQ), (PEC/PNEC), for pelagic organisms exposed to seepage of contaminated groundwater is therefore 5.0 μ g/L /6.7 μ g/L = 0.75. It is thus concluded that 1,2-dibromoethane-contaminated groundwater that is released to surface water is unlikely to cause direct adverse effects on populations of pelagic organisms in Canada.

In addition to the RQ calculated above for pelagic organisms, ChemSim model simulations were done to estimate the distance downstream from point of discharge where the acute and chronic threshold for 1,2-dibromoethaneis exceeded. These simulations considered three flow estimates (10th, 25th and 50th percentile) and two loading rates (0.1% with or without STP removal) as shown in Table 5 below. The 0.1% loading rate is calculated as 1% of the total release multiplied by the proportion of release to surface waters (10%).

In order to verify impacts of acute toxic effects of 1,2-dibromoethane, the lowest acceptable acute toxicity value was used in these simulations. Kszos et al. (2003) determined a 48-hour LC_{50} of 3.61 mg/L for *Ceriodaphnia dubia*. An application factor of 10 was used to account for species variability, to give a threshold for acute effects of 0.361 mg/L. This acute threshold is never exceeded along the plume centreline in any of the scenarios for more than 5 m downstream from the point of discharge.

For chronic effects, the PNEC for pelagic organisms (6.7 μ g/L) was used. Seven simulations were run (Table 5). The most conservative scenario (scenario 1) led to a situation where the chronic toxicity threshold is exceeded for a maximum of 755 m from the source along the plume centreline. However, it is considered very unlikely that this situation will occur because of the combination of worst-case assumptions, i.e., discharge

to a small river, low (10th percentile) flow and all of the substance used at one facility. For more realistic (less conservative, or protective) scenarios, this threshold is not exceeded for more than 100 m downstream from the point of discharge. Scenarios 6 and 7 are believed to be the most likely worst-case releases with low-flow conditions. For these, the concentrations of 1,2-dibromoethane at a distance of 50 m from the point of discharge are 3 and 2 μ g/L, respectively. The RQs, calculated as PEC/PNEC, for pelagic organisms exposed to surface waters are therefore 3 μ g/L/6.7 μ g/L = 0.45 and 2 μ g/L / 6.7 μ g/L = 0.30, respectively. Even at 10 m from the point of discharge, the RQs do not exceed 1. Consequently, the impacts seem very limited.

Table 5. ChemSim	modelling res	ults for the dist	ance downstr	eam from poi	int of
discharge where t	he PNEC for 1	,2-dibromoetha	aneis exceeded	l along the ce	ntreline of
the plume				-	

Run	Percentile flow	Release quantity divided among 8 facilities	STP removal	Release (kg/day)	PNEC exceeded (m)
1	10	no	no	0.075	755
2	10	no	yes	0.057	453
3	25	no	no	0.075	101
4	25	no	yes	0.057	53
5	50	no	no	0.075	13
6	10	yes	no	0.0094	8
7	10	yes	yes	0.0071	5

For soil organisms, the modelled LC_{50} value of 329.75 mg/L for the earthworm (ECOSAR 2008) was chosen as the CTV (with application of EqP equation to accommodate unit conversion from mg/L to mg/kg dw for soil), since reliable acute or chronic data were not identified. A PNEC of 3.3 mg/kg dw was derived by dividing the CTV by a factor of 10 to account for interspecies and intraspecies variations in sensitivities and by an additional factor of 10 to extrapolate from a modelled to an empirical value.

Two PEC values are used to characterize risk to soil organisms: one representative of industrial sites at 8×10^{-2} mg/kg dw and one for outside of non-industrial sites at 3.9×10^{-4} mg/kg dw.

The calculated RQs are 2.4×10^{-2} for industrial sites and 1.2×10^{-4} for non-industrial locations. It is therefore concluded based on the maximum measured concentrations in soil on industrial (as the worst-case scenario) as well as non-industrial sites that 1,2-dibromoethane is not likely to cause direct adverse effects on soil organisms in Canada. Furthermore, according to a report on environmental conditions at a chemical plant located in Ontario (Environment Canada 2001b), there is no evidence of contamination in either the shallow or deeper groundwater from monitored wells located in the periphery of the main chemical plant.

In summary, the risk quotient analysis indicates that 1,2-dibromoethane released to the Canadian environment is unlikely to cause adverse effects to pelagic and soil organisms.

Uncertainties in Evaluation of Ecological Risk

1,2-Dibromoethane is used as a scavenger of lead antiknock agents in leaded gasoline that is still used in Canada for some specific purposes, namely in piston engine aircrafts and competition vehicles. Based on its properties and empirical measurements, 1,2-dibromoethane is expected to be found in air, water and soil, but not in sediments. This substance has been found to be persistent in air and water, and has a high potential for long-range transport in air. It is not bioaccumulative. The confidence for the conclusions reached in this assessment is high. However, a few uncertainties are present and affect this assessment.

In the absence of measured values for releases of 1,2-dibromoethane other than for air, the proportions had to be estimated considering the uses and the chemistry of the substance. The estimated proportion of release to surface waters (10%) was used for simulations with ChemSim software.

For soil toxicity, no reliable empirical data have been found. In the absence of empirical data, a modelled QSAR value has been considered. Because of these uncertainties, conservative assumptions were made and high application factors were used.

No monitoring data for 1,2-dibromoethane have been found for soil and groundwater near storage tank systems in Canada. However, since presently in Canada there is limited approved use of leaded gasoline for piston engine aircraft (AvGas) and racing fuels for competition vehicles based on an exemption in the *Gasoline Regulations* under the *Canadian Environmental Protection Act, 1999* (CEPA 1999), the numbers of tanks containing 1,2-dibromoethane and that are potentially leaking should be limited.

Potential to Cause Harm to Human Health

Exposure Assessment

Environmental Media and Food

Empirical data were identified for environmental concentrations of 1,2-dibromoethane in raw and treated drinking water, soil, indoor air, ambient air, and food and beverages in Canada. Empirical data were also identified for environmental media in other locations. All studies identified containing empirical data for each environmental medium are summarized in Appendices 2–6.

In a recent study of contaminant levels in outdoor air conducted as part of the ongoing National Air Pollution Surveillance (NAPS) network, 1,2-dibromoethane was detected in only 7 of 1896 (or approximately 0.4%) samples, at a maximum concentration of 0.013

 μ g/m³, collected from 43 Canada-wide sites during the period January–December 2008 (NAPS 2008). The substance was detected near the method detection limit (0.025 μ g/m³) in a limited number of samples in outdoor air in Halifax in 2009 (Health Canada 2012). In 2007 in Regina, Saskatchewan and in 2005-2006 in Windsor, Ontario, 1,2-dibromoethane was not detected in outdoor air in either the summer or winter season (above the detection limits of 0.054 μ g/m³ and 0.123-0.15 μ g/m³, respectively) (Health Canada 2010b).

1,2-Dibromoethane was not detected in indoor air of residences in the summer and winter seasons of 2005 and 2006 in Windsor, Ontario, with detection limits of $0.123-0.15\mu g/m^3$ (Health Canada 2010b). In a 2007 study in Regina, Saskatchewan, the maximum 1,2-dibromoethane concentration in indoor air of $0.080 \ \mu g/m^3$ was measured in only one of 400 home samples at a detection limit of $0.054 \ \mu g/m^3$ (Health Canada 2010a). In addition, the substance was not detected in 643 indoor air samples in a study conducted in Halifax in 2009 above the detection limit of $0.025 \ \mu g/m^3$ (Health Canada 2012).

During the summer and winter seasons of 2005, a maximum concentration of 1,2-dibromoethane in personal breathing-zone air of 0.190 μ g/m³ was measured in Windsor, Ontario (Health Canada 2010b). Participants wore padded backpacks with samplers that provided concentrations of selected volatile organic compounds averaged over a 24-hour period for five consecutive days (Health Canada 2010b). Less than 0.5% of the samples were above the detection limit of 0.123 μ g/m³.

In an expansive survey of raw, treated and distribution water in Ontario collected between January 1, 2005, and December 31, 2006, 1,2-dibromoethane was not detected in any sample (detection limit = $0.1 \ \mu g/L$) (Ontario MOE 2006). Other Canadian studies conducted between 2002 and 2008 did not detect 1,2-dibromoethane in drinking water (City of Victoria 2008; Ville de Montréal 2006; NSEL 2005; COWQS 2003). A summary of drinking water data obtained from sites distributed across the United States provided by the United States Geological Survey over a sampling period of 1985–2001 revealed median 1,2-dibromoethane levels of "< $0.10 \ \mu g/L$ " and "< $0.04 \ \mu g/L$ " for public wells and domestic wells, respectively (Zogorski et al. 2006).

Data on 1,2-dibromoethane concentrations in food in Canada were identified. Ten flour samples in 1984 in Saskatoon, Saskatchewan, contained a maximum concentration of 405.3 μ g/kg (McKay 1986). However, 1,2-dibromoethane was discontinued as an agricultural pesticide in Canada in 1984 (UNEP and FAO 2003), and there have been no reports of concentrations in cereals or cereal products since then in Canada. Residuals of 1,2-dibromoethane in foodstuffs are not currently monitored by the Canadian Food Inspection Agency (August 2009 email from Canadian Food Inspection Agency to Existing Substances Risk Assessment Bureau, Health Canada; unreferenced). In general, processing, cooking, baking and market circulation of food items decrease residual levels of 1,2-dibromoethane (Konishi et al. 1986). The ban on the use of 1,2-dibromoethane as an agricultural pesticide in North America and Europe (UNEP and FAO 2003) has reduced the likelihood of exposure of the Canadian population to the chemical in domestic and imported food. While a few countries that still apply 1,2-dibromoethane to

foodstuffs have been identified—namely, five countries identified in a search of 90 countries in the Homologa database (Tanzania, South Africa, India, Zimbabwe and Zambia) and some honey producers in Greece (Tananaki et al. 2005, 2006), the contribution of these foods to the Canadian food supply is considered minimal. As the majority of food studies identified had sampling periods in the 1980s when substantial global use of 1,2-dibromoethane as a pesticide may have still occurred, the data are not considered applicable to the current context.

A study released by the United States Food and Drug Administration (US FDA) in 2003 identified a 1,2-dibromoethane concentration of 13 μ g/kg in one sample of imported sweet cucumber pickle (US FDA 2003). In studies in Greece with sampling periods from 2003 to 2005, the maximum concentration of 1,2-dibromoethane in bulk honey was 331.2 μ g/kg (Tananaki et al. 2005, 2006). Despite a recommendation by the Greek Hellenic Food Authority to beekeepers to abandon use of 1,2-dibromoethane as a moth control agent, some individual beekeepers may still use it (Tananaki et al. 2006).

In a 1993 Canadian study, the maximum 1,2-dibromoethane concentrations detected in urban parkland and rural parkland soils in Ontario in the early 1990s were 0.032 ng/g and 0.390 ng/g, respectively (OMEE 1993). 1,2-Dibromoethane was discontinued as an agricultural pesticide in Canada in 1984.

Based on the current use pattern of 1,2-dibromoethane and the recent Canadian monitoring data, particularly for air, exposure to the general population is expected to be low to negligible.

Consumer Products

No consumer products containing 1,2-dibromoethane were reported in responses to the section 71 survey issued under CEPA 1999 (Environment Canada 2001a), and no data were identified on exposure to 1,2-dibromoethane through use of consumer products. Therefore, exposure to 1,2-dibromoethane from use of consumer products is not expected.

Confidence in Exposure Assessment

Confidence in the environmental exposure database is considered to be moderate while for the food database, it is considered to be low. Empirical data were obtained for all environmental media, and the data were specific to Canada; however, the information for foodstuffs was not specific to Canada. Given current use patterns of the substance in Canada and internationally, and on the sporadic detection of 1,2-dibromoethane in monitoring studies, there is confidence that actual exposures to the general population are low to negligible.

Health Effects Assessment

Appendix 7 provides an overview of the health effects information for 1,2-dibromoethane.

The International Programme on Chemical Safety (IPCS 1996) concluded that 1,2-dibromoethane is a carcinogen in rodents and a potential human carcinogen. The International Agency for Research on Cancer (IARC 1999) concluded that there is inadequate evidence in humans and sufficient evidence in experimental animals for the carcinogenicity of 1,2-dibromoethane; 1,2-dibromoethane was classified as probably carcinogenic to humans (Group 2A). In addition, it is classified in the European Union as Carcinogen Category 1B, with the hazard statement "May cause cancer" (European Union 2008). These conclusions were based on significant increases in tumour incidences in rats and mice exposed via multiple routes. Significant increases in the incidences of squamous cell carcinomas of the forestomach were observed in male and female rats administered 1,2-dibromoethane by gavage at 37 mg/kg-bw per day or more for up to 61 weeks (NCI 1978). Rats were exposed by inhalation to 0, 10, or 40 ppm (equivalent to 0, 77 or 308 mg/m³) for 88–103 weeks. There were significant increases in the incidence of nasal cavity carcinomas at high doses (males: controls, 0/50; high dose, 21/50; females: controls, 0/50; high dose, 25/50), adenocarcinomas at both doses (males: controls, 0/50; low dose, 20/50; high dose, 28/50; females: controls, 0/50; low dose, 20/50; high dose, 29/50) and adenomas at low doses (males: control, 0/50; low dose, 11/50; females: controls, 0/50; low dose, 11/50). In addition, significant increases in the incidences of mammary gland fibroadenomas in females and tunica vaginalis mesotheliomas and nasal cavity adenomatous polyps in males were reported (NTP 1982). Dermal exposure of mice to 25 or 50 mg/day (equivalent to 357 or 714 mg/kg-bw per day, respectively; as per Health Canada 1994) for up to 594 days resulted in an increased incidence of papillomas of the lungs in female mice (and a significant increase in the incidence of skin papillomas and carcinomas at 50 mg/day) (Van Duuren et al. 1979). In each of these bioassays, these significant increases were observed at the lowest exposure level tested and higher.

1,2-Dibromoethane was genotoxic in a large number and variety of assays, including *in vivo* deoxyribonucleic acid (DNA) binding and DNA damage in mice and rats and mammalian cell mutagenicity assays and *in vitro* mutagenicity, clastogenicity and DNA damage assays (see Appendix 7).

The United States Environmental Protection Agency (US EPA) has derived cancer potency estimates for 1,2-dibromoethane. A cancer oral slope factor of 1.8 (mg/kg bw/day)⁻¹ was derived, based on the incidence of forestomach tumours in male rats in the oral carcinogenicity study mentioned above, but this "...factor should not be used with exposures greater than approximately 0.5 mg/kg/day, since the observed dose-response would not be expected to continue linearly above this estimated lifetime-equivalent exposure level." An inhalation cancer slope factor of 0.6 (mg/m³)⁻¹ was estimated based on the incidence of nasal cavity tumours in male rats in the inhalation carcinogenicity study mentioned above, but this "...unit risk should not be used with exposures greater than

 0.023 mg/m^3 (0.18 ppm), because above this level, the dose-response is not linear." (see US EPA 2004). Several uncertainties limit the confidence in use and derivation of these slope factors, such as the high mortality limiting the duration of the study and close dose spacing in the oral rat study, and high mortality in both rats and mice, especially in the high-dose groups, in the inhalation carcinogenicity studies (see Appendix 7).

Cancer potency factors were also derived by Health Canada. A lowest tumorigenic dose $05 (TD_{05})$ of 0.04 mg/kg-bw per day was calculated, based on the incidence of squamous cell carcinomas in the forestomach of male rats in the oral carcinogenicity study mentioned above. The TD₀₅ is defined as the total intake associated with a 5% increase in incidence or mortality due to tumours scaled, where appropriate, to reflect interspecies variations. Although the exposure levels and the overall duration of the oral rat study were reduced due to excessive mortality, note that the derived TD₀₅ is based on the low doses. A highest tumorigenic concentration 05 (TC₀₅) was not calculated due to the high mortality in both rats and mice, especially in the high-dose groups, in the inhalation carcinogenicity studies mentioned above. The TC₀₅ is defined as the concentration, generally in air, associated with a 5% increase in incidence or mortality due to tumours (Health Canada 1996).

Male reproductive effects are considered to be the critical non-cancer effect. In a short-term longitudinal study in male forestry workers (engaged in the application or spraying of 1,2-dibromoethane [4% by volume] emulsion), significantly decreased sperm velocity and semen volume were observed in subjects exposed via inhalation to 1.2-dibromoethane levels of 0.46 mg/m³ or greater (occupational time-weighted average) in conjunction with dermal exposure (Schrader et al. 1988; also cited in IPCS 1996). The authors did not report exposure to any other chemicals in the forestry workers engaged in the application or spraying activities. Longer-term exposure to 1,2-dibromoethane ranging from mean concentration of 88 ppb to peak exposure of up to 262 ppb (0.68 to 2.0 mg/m^3 ; IPCS 1996) in the fumigation workers caused significant reductions in sperm count, viable sperms and increase in number of abnormal sperms (Ratcliffe et al. 1987). Male reproductive effects were also observed in multiple species of experimental animals exposed to the lowest doses or concentrations tested and higher. The lowest lowest-observed-effect level (LOEL) for reproductive effects for the oral route of exposure was 2 mg/kg-bw per day based on reversible low sperm density, poor motility and altered spermatozoa morphology observed in a 2-year study in bulls (Amir and Volcani 1965). In another study, testicular atrophy was seen in male rats following long-term oral exposure (38 mg/kg-bw per day) to 1,2-dibromoethane (NCI 1978). Testicular degeneration in male rats was observed at an inhalation concentration of 77 mg/m³ in conjunction with other non-cancer effects, including toxic nephropathy in males, retinal atrophy and adrenal cortex degeneration in females and increases in hepatic necrosis in both sexes (NTP 1982). Similarly, the reproductive effects were reported in male or female rats following inhalation exposure to 89 or 80 ppm (equivalent to 684 or 614 mg/m³ as per IPCS 1996) for 10 or 3 weeks, respectively. Effects in male rats included reduction in testicular weight: decreased serum testosterone levels: atrophy of testes, epididymis, prostate and seminal vesicles; and changes in reproductive behaviour.

Female rats exposed to 1,2-dibromoethane had altered estrous cycle until several days after cessation of exposure (Short et al. 1979).

Limited information was available regarding the acute effects of 1,2-dibromoethane in humans. A review of 64 cases of acute poisoning in humans reported that ingestion of 1.5 ml (estimated to be more than 3000 mg) of 1,2-dibromoethane may be fatal in humans. Effects observed included nausea, vomiting, abdominal pain and signs of hepatotoxicity, nephrotoxicity, nervous system toxicity and cardiotoxicity in male and female patients (Singh et al. 2007).

Characterization of Risk to Human Health

General population exposure to 1,2-dibromoethane is expected to be low to negligible from air based on the specialized use pattern of the substance and as the substance has not been detected (> 99% of the time) at low levels in recently conducted monitoring studies of outdoor and indoor air and personal air. As no consumer products containing 1,2-dibromoethane had been identified in Canada, consumer product exposure is not expected.

A critical effect for the characterization of risk of 1,2-dibromoethane exposure to human health is carcinogenicity, as there is evidence of carcinogenicity of 1,2-dibromoethane in rats and mice following oral or inhalation exposure. Moreover, the positive genotoxicity results reported in several *in vivo* and *in vitro* studies suggest that the potential for 1,2-dibromoethane to induce tumours through direct interaction with genetic material cannot be precluded.

On the basis of the use pattern of 1,2-dibromethane and the very limited potential for general population exposure, it is concluded that 1,2-dibromoethane is not a substance that is entering the environment in a quantity or concentration or under conditions that constitute or may constitute a danger in Canada to human life or health.

Uncertainties in Evaluation of Risk to Human Health

Based on the extensive dataset on carcinogenicity and *in vivo* and *in vitro* genotoxicity assays, there is high confidence in the conclusion that 1,2-dibromoethane is considered to induce tumours through direct interaction with genetic material. However, uncertainties exist regarding inter- and intraspecies variation, extrapolation of data from animals to humans and lack of data in humans for several endpoints. Based on the use pattern and extensive monitoring of 1,2-dibromoethane in outdoor and indoor air, there is confidence that exposure to the general population is low to negligible. Targeted ambient air monitoring of 1,2-dibromoethane in the vicinity of sites of its known use would reduce any remaining uncertainty in this conclusion.

Conclusion

Based on the information available with regard to the environment, it is concluded that 1,2-dibromoethane is not entering the environment in a quantity or concentration or under conditions that have or may have an immediate or long-term harmful effect on the environment or its biological diversity or that constitute or may constitute a danger to the environment on which life depends. Additionally, it is concluded that 1,2-dibromoethane meets the criteria for persistence but not for bioaccumulation potential as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

On the basis of the use pattern of 1,2-dibromethane and the very limited potential for general population exposure, it is concluded that 1,2-dibromoethane is not entering the environment in a quantity or concentration or under conditions that constitute or may constitute a danger in Canada to human life or health.

It is therefore concluded that 1,2-dibromoethane does not meet one or more criteria under section 64 of CEPA 1999.

Because this substance is listed on the Domestic Substances List, its import and manufacture in Canada are not subject to notification under subsection 81(1). Given the hazardous properties of this substance, there is concern that new activities that have not been identified or assessed could lead to this substance meeting the criteria set out in section 64 of the Act. Therefore, it is recommended to amend the Domestic Substances List, under subsection 87(3) of the Act, to indicate that subsection 81(3) of the Act applies with respect to the substance so that new manufacture, import or use of this substance is notified and undergoes ecological and human health risk assessments.

References

Alexeeff GV, Kilgore WW, Li MY. 1990. Ethylene dibromide: toxicology and risk assessment. New York (NY): Springer-Verlag. p. 49–122 [cited in IPCS 1996].

Alper MD, Ames BN. 1975. Positive selection of mutants with deletions of the *gal-ch1* region of the *Salmonella* chromosome as a screening procedure for mutagens that cause deletions. J Bacteriol 121:259–266 [cited in IPCS 1996].

Ames BN, Yanofsky C. 1971. The detection of chemical mutagens with enteric bacteria. In: Hollaender A, editor. Chemical mutagens: principles and methods for their detection, vol. 1. New York (NY): Plenum Press. p. 267–282 [cited in IPCS 1996].

Amir D, Volcani R. 1965. Effect of dietary ethylene dibromide on bull semen. Nature 206:99–100.

Arfellini G, Bartoli S, Colacci A, Mazzullo M, Galli MC, Prodi G, Grilli S. 1984. *In vivo* and *in vitro* binding of 1,2-dibromoethane and 1,2-dichloroethane to macromolecules in rat and mouse organs. J Cancer Res Clin Oncol 108:204–213 [cited in IARC 1999].

Asita AO, Hayashi M, Kodama Y, Matsuoka A, Suzuki T, Sofuni T. 1992. Micronucleated reticulocyte induction by ethylating agents in mice. Mutat Res 271:29–37 [cited in IPCS 1996; IARC 1999].

[ATSDR] Agency for Toxic Substances and Disease Registry. 1992. Toxicological profile for 1,2-dibromoethane. Atlanta (GA): Public Health Service, US Department of Health and Human Services.

Ballering LAP, Nivard MJM, Vogel EW. 1993. Characterization of the genotoxic action of three structurally related 1,2-dihaloalkanes in *Drosophila melanogaster*. Mutat Res 285:209–217 [cited in IPCS 1996; IARC 1999].

Ballering LAP, Nivard MJM, Vogel EW. 1994. Mutation spectra of 1,2-dibromoethane, 1,2-dichloroethane and 1-bromo-2-chloroethane in excision repair proficient and repair deficient strains of *Drosophila melanogaster*. Carcinogenesis 15:869–875 [cited in IARC 1999].

Barber ED, Donish WH. 1982. An exposure system for quantitative measurements of the microbial mutagenicity of volatile liquids. Measurements of microbial mutagenicity. In: Tice RR, Costa DL, Schaich KM, editors. Genotoxic effects of airborne agents. New York (NY): Plenum Press. p. 3–18 [cited in IPCS 1996].

Barber ED, Donish WH, Mueller KR. 1981. A procedure for the quantitative measurement of the mutagenicity of volatile liquids in the Ames *Salmonella*/microsome assay. Mutat Res 90:31–48 [cited in IPCS 1996; IARC 1999].

Barnett LB, Lovell DP, Felton CF, Gibson BJ, Cobb RR, Sharpe DS, Shelby MD, Lewis SE. 1992. Ethylene dibromide: negative results with the mouse dominant lethal assay and the electrophoretic specific locus test. Mutat Res 282:127–133 [cited in IPCS 1996; IARC 1999].

Bentley KS, Working PK. 1988. Activity of germ-cell mutagens and nonmutagens in the rat spermatocyte UDS assay. Mutat Res 282:127–133 [cited in IARC 1999].

Beyer A, Mackay D, Matthies M, Wania F, Webster E. 2000. Assessing long-range transport potential of persistent organic pollutants. Environ Sci Technol 34(4):699–703.

Bjørge C, Brunborg G, Wiger R, Holme JA, Scholz T, Dybing E, Søderlund EJ. 1996. A comparative study of chemically induced DNA damage in isolated and human and rat testicular cells. Reprod Toxicol 10:509–519.

Bradley MO, Dysart G. 1985. DNA single-strand breaks, double-strand breaks, and cross-links in rat testicular germ cells: measurements of their formation and repair by alkaline and neutral filter elution. Cell Biol Toxicol 1:341–346 [cited in IARC 1999].

Brem H, Stein AB, Rosenkranz HS. 1974. The mutagenicity and DNA-modifying effect of haloalkanes. Cancer Res 34:2576–2579 [cited in IPCS 1996; IARC 1999].

Brimer PA, Tan E-L, Hsie AW. 1982. Effect of metabolic activation on the cytotoxicity and mutagenicity of 1,2-dibromoethane in the CHO/HGPRT system. Mutat Res 95:377–388 [cited in IPCS 1996; IARC 1999].

Brown SK, Sim MR, Abramson MJ, Gray CN. 1994. Concentrations of volatile organic compounds in indoor air—a review. Indoor Air 4:123–134.

Budavari S, O'Neil MJ, Smith A, Heckelman PE, editors. 2001. The Merck index—an encyclopedia of chemicals, drugs and biologicals. Whitehouse Station (NJ): Merck Research Laboratories, Division of Merck & Co. Inc. p. 675.

Buijs W, van der Gen A, Mohn GR, Breimer DD. 1984. The direct mutagenic activity of alpha, omega-dihalogenoalkanes in *Salmonella typhimurium*. Strong correlation between chemical properties and mutagenic activity. Mutat Res 141:11–14 [cited in IPCS 1996].

[Cal EPA] California Environmental Protection Agency. 1992. Indoor pollutant concentrations and exposures. Contract No. A833-156. Sacramento (CA): California Environmental Protection Agency, Air Resources Board.

Canada. [1978]. *Food and Drug Regulations*, C.R.C., c. 870. Available from: <u>http://laws.justice.gc.ca/en/showtdm/cr/C.R.C.-c.870</u> Canada. 1990. *Canadian Environmental Protection Act, 1999: Gasoline Regulations,* P.C. 1990-740, 26 April 1990, SOR/90-247. Canada Gazette, Part II, vol. 124, no. 10. Available from: <u>http://laws.justice.gc.ca/PDF/Regulation/S/SOR-90-247.pdf</u>

Canada. 1999. *Canadian Environmental Protection Act, 1999.* S.C., 1999, c. 33. Canada Gazette, Part III, vol 22, no. 3. Available from: http://www.gazette.gc.ca/archives/p3/1999/g3-02203.pdf

Canada. 2000. *Canadian Environmental Protection Act: Persistence and Bioaccumulation Regulations*, P.C. 2000-348, 23 March 2000, SOR/2000-107. Canada Gazette, Part II, vol. 134, no. 7. p. 607–612. Available from: http://www.gazette.gc.ca/archives/p2/2000/2000-03-29/pdf/g2-13407.pdf

Canada, Dept. of the Environment. 2001. *Canadian Environmental Protection Act, 1999: Notice with respect to certain substances on the Domestic Substances List (DSL)*. Canada Gazette, Part I, vol. 135, no. 46, p. 4194–4211. Available from: <u>http://www.gazette.gc.ca/archives/p1/2001/2001-11-17/pdf/g1-13546.pdf</u>

Canada. 2010. *Canadian Environmental Protection Act, 1999: Regulations Amending the Gasoline Regulations*, P.C. 2101-796, 17 June 2010, SOR/2010-134. Canada Gazette, Part II, vol. 144, no. 14. Available from: <u>http://www.gazette.gc.ca/rp-pr/p2/2010/2010-07-07/pdf/g2-14414.pdf#page=2</u>

Channarayappa, Ong T, Nath J. 1992. Cytogenetic effects of vincristine sulfate and ethylene dibromide in human peripheral lymphocytes: micronucleus analysis. Environ Mol Mutagen 20:117–126 [cited in IPCS 1996; IARC 1999].

ChemSim. 2003. Chemical Release and Dispersion Analysis Application. Version 2.0.5. Developed by Canadian Hydraulics Centre, National Research Council of Canada, Ottawa (ON).

Chroust K, Pavlova M, Prokop Z, Mendel J, Bozkova K, Kubat Z, Zajickova V, Damborsky J. 2007.Quantitative structure-activity relationships for toxicity and genotoxicity or halogenated aliphatic compounds: wing spot test of *Drosophila melanogaster*. Chemosphere 67(1):152–159.

[CITI] Chemical Inspection and Testing Institute. 1992. Biodegradation and Bioaccumulation Data of Existing Chemicals Based on the CSCL Japan. Japan Chemical Industry Ecology-Toxicology & Information Center. p. 2–14.

City of Toronto. 1990. The quality of drinking water in Toronto: a review of tap water, bottled water and water treated by a point-of-use device. Toronto (ON): City of Toronto, Department of Public Health. 133 p.

City of Victoria. 2008. 2008 untreated (raw) water quality at Japan Gulch Plant. Victoria (BC): Capital Regional District Water Services.

Class TH, Ballschmiter K. 1988. Chemistry of organic traces in air. VIII. Sources and distribution of bromo- and bromochloromethanes in marine air and surface water of the Atlantic Ocean. J Atmos Chem 7:35–46.

Clive D, Johnson KO, Spector JFS, Batson AG, Brown MMM. 1979. Validation and characterization of the L5178Y/TK+/– mouse lymphoma mutagen assay system. Mutat Res 59:61–108 [cited in IPCS 1996; IARC 1999].

Clower M Jr, McCarthy JP, Rains DM. 1985. Effect of cooking on levels of ethylene dibromide residues in rice. J Assoc Off Anal Chem 68(4):710–711.

Clower M Jr, McCarthy JP, Carson LJ. 1986. Comparison of methodology for determination of ethylene dibromide in grains and grain-based foods. J Assoc Off Anal Chem 69(1):87–90.

Cmarik JL, Inskeep PB, Meredith MJ, Meyer DJ, Ketterer B, Guengerich FP. 1990. Selectivity of rat and human glutathione *S*-transferases in activation of ethylene dibromide by glutathione conjugation and DNA binding and induction of unscheduled DNA synthesis in human hepatocytes. Cancer Res 50:2747–2752 [cited in IPCS 1996; IARC 1999].

[CMHC] Canada Mortgage and Housing Corporation. 1989. CHMC Kitchener townhouse study of soil gas ventilation as a remedial measure for methane entry into basements. Prepared for Research Division, Canada Mortgage and Housing Corporation, Ottawa, Ontario. Waterloo (ON): CH2M Hill Engineering Ltd.

[CNS] Cosmetic Notification System [Proprietary Database]. 2009. [cited 2009 Aug 5]. Available from Health Canada, Cosmetics Division.

Cohen MA, Ryan PB, Yanagisawa Y, Spengler JD, Özkaynak H, Epstein PS. 1989. Indoor–outdoor measurements of volatile organic compounds in the Kanawha Valley of West Virginia. J Air Pollut Control Assoc 39:1086–1093.

Colacci A, Mazzulo M, Arfellini G, Prodi G, Grilli S. 1995. *In vitro* microsome- and cytosol-mediated binding of 1,2-dichloroethane and 1,2-dibromoethane with DNA. Cell Biol Toxicol 1:45–55 [cited in IARC 1999].

[COWQS] City of Ottawa Water Quality Section. 2003. 2003 Organics Summary. Ottawa (ON): City of Ottawa.

Crebelli R, Conti G, Conti L, Carere A. 1984. Induction of somatic segregation by halogenated aliphatic hydrocarbons in *Aspergillus nidulans*. Mutat Res 138:33–38 [cited in IPCS 1996].

Crespi CL, Seixas GM, Turner TR, Ryan CG, Penman BW. 1985. Mutagenicity of 1,2-dichloroethane and 1,2-dibromoethane in two human lymphoblastoid cell lines. Mutat Res 142:133–140 [cited in IPCS 1996; IARC 1999].

Daft JL. 1988. Rapid determination of fumigant and industrial chemical residues in food. J Assoc Off Anal Chem 71(4):748–760.

DeSerres FJ, Malling HV. 1983. The role of *Neurospora* in evaluating environmental chemicals for mutagenic activity. Ann N Y Acad Sci 407:177–185 [cited in IPCS 1996].

Dobbins JG. 1987. Regulation and the use of "negative" results from human reproductive studies: the case of ethylene dibromide. Am J Ind Med 12:33–45.

Dunkel VC, Zeiger E, Brusick D, McCoy E, McGregor D, Mortelmans K, Rosenkranz HS, Simmon VF. 1985. Reproducibility of microbial mutagenicity assays: testing of carcinogens and noncarcinogens in *Salmonella typhimurium* and *Escherichia coli*. Environ Mutagen 7:1–248 [cited in IPCS 1996; IARC 1999].

[ECOSAR] Ecological Structural Activity Relationships [Internet]. 2008. Version 1.00. Washington (DC): US Environmental Protection Agency, Office of Pollution Prevention and Toxics; Syracuse (NY): Syracuse Research Corporation. Available from: http://www.epa.gov/oppt/exposure/pubs/episuite.htm.

Elliott BM, Ashby J. 1980. Ethylene dibromide and disulfiram: studies *in vivo* and *in vitro* on the mechanism of the observed synergistic carcinogenic response. Carcinogenesis 1:1049–1057 [cited in IPCS 1996; IARC 1999].

Environment Canada. 1991. Measurement program for toxic contaminants in Canadian urban air: update and summary report. Ottawa (ON): Environment Canada, Environmental Technology Centre. Report Series No. PMD 91-2.

Environment Canada. 1992. Detroit incinerator monitoring program: data report #6. Ottawa (ON): Environment Canada, Environmental Technology Centre, Pollution Measurement Division. Report Series No.92-1.

Environment Canada. 1994. Volatile organic compound measurements in the Greater Vancouver Regional District(GVRD) 1989–1992. Ottawa (ON): Environment Canada, Environmental Technology Centre, Pollution Measurement Division. Report Series No. PMD 94-1.

Environment Canada. 1995. Volatile organic compounds in the ambient air of the province of Quebec (1989–1993). Quebec City (QC): Environment Canada, Atmospheric Pollution and Toxic Substances Control Division and Pollution Measurement Division, Quebec Region.

Environment Canada. 1996. Ecological effects assessments of priority substances under the *Canadian Environmental Protection Act*. Resource Document. Draft 2.0. Gatineau (QC): Environment Canada, Commercial Chemicals Evaluation Branch, Chemicals Evaluation Division. Section 6-19.

Environment Canada. 2001a. Data collected under the *Canadian Environmental Protection Act, 1999,* Section 71: *Notice with respect to certain substances on the Domestic Substances List.* Data prepared by: Environment Canada, Existing Substances Program.

Environment Canada. 2001b. Data collected under the *Canadian Environmental Protection Act, 1999,* Section 71: *Notice with respect to certain substances on the Domestic Substances List.* Data prepared by: Environment Canada, Existing Substances Program. Study Submission 2001.

Environment Canada. 2004. Measured ethylene dibromide levels in air in Canada. Provided by the Environmental Science and Technology Centre, Analysis and Air Quality Division, Environment Canada, Ottawa (ON).

Environment Canada. 2009a. Environment Canada's *Gasoline Regulations*. Discussion paper proposing indeterminate exemption for the use of leaded gasoline in competition vehicles. December 2009. Available from: http://www.ec.gc.ca/CEPARegistry/documents/regs/leaded_gasoline/index.cfm

Environment Canada. 2009b. Measured ethylene dibromide levels in air in Canada. Provided by the Environmental Science and Technology Centre, Analysis and Air Quality Division, Environment Canada, Ottawa (ON).

Epstein SS, Arnold E, Andrea J, Bass W, Bishop Y. 1972. Detection of chemical mutagens by the dominant lethal assay in the mouse. Toxicol Appl Pharmacol 23:288–325 [cited in IPCS 1996; IARC 1999].

[EQC] Equilibrium Criterion Model. 2003. Version 2.02. Peterborough (ON): Trent University, Canadian Environmental Modelling Centre.Available from: http://www.trentu.ca/academic/aminss/envmodel/models/EQC2.html

European Union. 2008. REGULATION (EC) No 1272/2008 OF THE EUROPEAN PARLIAMENT AND OF THE COUNCIL of 16 December 2008 on classification, labelling and packaging of substances and mixtures, amending and repealing Directives 67/548/EEC and 1999/45/EC, and amending Regulation (EC) No 1907/2006. Official Journal of the European Union L 353. 31 December 2008. Available at: <u>http://eurlex.europa.eu/LexUriServ/LexUriServ.do?uri=OJ:L:2008:353:0001:1355:EN:PDF</u>

Fahrig R. 1974. Comparative mutagenicity studies with pesticides. In: Chemical carcinogenesis essays. Lyon (FR): International Agency for Research on Cancer. p. 161–181. IARC Scientific Publications No. 10 [cited in IPCS 1996].

Falta RW, Bulsara N, Henderson JK, Mayer RA. 2005. Leaded-gasoline additives still contaminate groundwater. Environ Sci Technol 39(18):379–384.

Ferreri AM, Rocchi P, Capucci A, Prodi G. 1983. Induction of diphtheria toxin–resistant mutants in human cells by halogenated compounds. J Cancer Res Clin Oncol 105:111–112 [cited in IPCS 1996; IARC 1999].

Foster PL, Wilkinson WG, Miller JK, Sullivan AD, Barnes WM. 1988. An analysis of the mutagenicity of 1,2-dibromoethane to *Escherichia coli*: influence of DNA repair activities and metabolic pathways. Mutat Res 194:171–181 [cited in IPCS 1996].

Foureman P, Mason JM, Valencia R, Zimmering S. 1994. Chemical mutagenesis testing in *Drosophila*. X. Results of 70 coded chemicals tested for the National Toxicology Program. Environ Mol Mutagen 23:208–227 [cited in IARC 1999].

Gerhartz W, executive editor. 1985. Ullmann's encyclopedia of industrial chemistry, vol. A4. 5th ed. Deerfield Beach (FL): VCH Publishers. p. 409.

Going JE, Long S. 1975. Sampling and analysis of selected toxic substances: Task II — Ethylene dibromide. Washington (DC): Office of Toxic Substances, US Environmental Protection Agency. (EPA-560/6-75-001).

Golder Associates. 1987. Testing of specific organic compounds in soils in background in urban areas; Port Credit and Oakville/Burlington, ON. Draft working paper to Shell Canada Ltd. and Texaco Canada Ltd., Mississauga (ON).

Graf U, Wurgler FE, Katz AJ, Frei H, Juon H, Hall CB, Kale PG. 1984. Somatic mutation and recombination test in *Drosophila melanogaster*. Environ Mutagen 6:153–188 [cited in IPCS 1996].

Great Lakes Commission. 2002. Inventory of Toxic Air Emissions — December 2002. Ontario Toxic Emission Inventory, Great Lakes Commission.

Gunderson EL. 1988a. FDA total diet study, April 1982 – April 1986, dietary intakes of pesticides, selected elements, and other chemicals. Data tables. Washington (DC): US Food and Drug Administration, Division of Contaminants Chemistry. Study details available from: <u>http://www.cfsan.fda.gov/~comm/tds-toc.html</u>

Gunderson EL. 1988b. FDA total diet study, April 1982 – April 1984, dietary intake of pesticides, selected elements, and other chemicals. J Assoc Off Anal Chem 71(6):1200–1209.

Hawkins WE, Walker WW, James MO, Manning CS, Barnes DH, Heard CS, Overstreet RM. 1998. Carcinogenic effects of 1,2-dibromoethane (ethylene dibromide; EDB) in Japanese medaka (*Oryzias latipes*). Mutat Res 399(2):221-32.

Hayes S, Gordon A, Sadowski I, Hayes C. 1984. RK bacterial test for independently measuring chemical toxicity and mutagenicity: short-term forward selection assay. Mutat Res 130:97–106 [cited in IPCS 1996].

Health Canada. 1994. Human health risk assessment for priority substances. Ottawa (ON): Health Canada. Available from: <u>http://www.hc-sc.gc.ca/ewh-semt/alt_formats/hecs-sesc/pdf/pubs/contaminants/approach/approach-eng.pdf</u>

Health Canada. 1996. Health-based tolerable daily intakes/concentrations and tumorigenic doses/concentrations for priority substances. Cat. H46-2/96-194E. Minister of Supply and Services Canada 1996.

Health Canada. 2004. State of the science report for a screening health assessment: 1,2-dibromoethane. Ottawa (ON): Health Canada. Available from: <u>http://www.hc-sc.gc.ca/ewh-semt/alt_formats/hecs-sesc/pdf/pubs/contaminants/dibromoethane/dibromoethane-eng.pdf</u>

Health Canada. 2010a. Regina Indoor Air Quality Study (2007):Data Summary for Volatile Organic Compound Sampling. Cat.: H128-1/10-617E-PDF. Ottawa (ON): Health Canada 2010.

Health Canada. 2010b. Windsor Exposure Assessment Study (2005–2006): Data Summary for Volatile Organic Compound Sampling. Cat.: H128-1/10-618E-PDF Ottawa (ON): Health Canada 2010.

Health Canada. 2012. Halifax Indoor Air Quality Study (2009) - Volatile Organic Compounds (VOC) Data Summary. Cat.: H129-19/2012E-PDF. Ottawa(ON): Health Canada 2012.

Hemminki K, Falck K, Vainio H. 1980. Comparison of alkylation rates and mutagenicity of directly acting industrial and laboratory chemicals. Arch Toxicol 46:277–285 [cited in IPCS 1996].

Holcombe GW, Benoit DA, Hammermeister DE, Leonard EN, Johnson RD. 1995. Acute and long-term effects of nine chemicals on the Japanese medaka (*Oryzias latipes*). Arch Environ Contam Toxicol 28:287–297.

Holzer J, Voss B, Karroum S, Hildmann H, Wilhelm M. 2008. A comparative study of chemically induced DNA damage in isolated nasal mucosa cells of humans and rats assessed by the alkaline comet assay. J Toxicol Environ Health A 71:936–946.

Howard PH, Boethling RS, Jarvis WF, Meylan WM, Michalenko EM. 1991. Handbook of environmental degradation rates. 2nd ed. Chelsea (MI): Lewis Publishers. p. 378–379.

[HSDB] Hazardous Substances Data Bank [database on the Internet]. 1983–2010. Bethesda (MD): US National Library of Medicine. [revised 2005 June 24; cited 2010 Feb. 9]. Available from: <u>http://toxnet.nlm.nih.gov/cgi-bin/sis/htmlgen?HSDB</u>

Hughes TJ, Simmons DM, Monteith LG, Claxton LD. 1987. Vaporization technique to measure mutagenic activity of volatile organic chemicals in the Ames/*Salmonella* assay. Environ Mutagen 9:421–441 [cited in IPCS 1996].

[IARC] International Agency for Research on Cancer. 1999. Re-evaluation of some organic chemicals, hydrazine and hydrogen peroxide (Part 2). IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. IARC Monogr Eval Carcinog Risks Hum 71:641–669.

Inskeep PB, Koga N, Cmarik JL, Guengerich FP. 1986. Covalent binding of 1,2-dihaloalkanes to DNA and stability of the major DNA adduct, *S*-[2-(N7-guanyl)ethyl]glutathione. Cancer Res 46:2839–2844 [cited in IPCS 1996; IARC 1999].

[IPCS] International Programme on Chemical Safety. 1996. 1,2-Dibromoethane. Geneva (CH): World Health Organization. (Environmental Health Criteria 177). Jointly sponsored by the United Nations Environment Programme, the International Labour Organization and the World Health Organization.

[IRIS]. Integrated Risk Information System Database. 2002. 1,2-Dibromoethane (CASRN 106-93-4). Washington (DC): Environmental Criteria and Assessment Office, US Environmental Protection Agency. [cited 2003 October]. Available from: http://cfpub.epa.gov/iris/quickview.cfm?substance nmbr=0361

[IUCLID] International Uniform Chemical Information Database. 2000. IUCLID data sheet: 1,2-Dibromoethane. Substance ID: 106-93-4. International Uniform Chemical Information Database, European Chemicals Bureau, European Commission.

Ivett JL, Brown BM, Rodgers C, Anderson BE, Resnick MA, Zeiger E. 1989. Chromosomal aberrations and sister chromatid exchange tests in Chinese hamster ovary cells. IV. Results with 15 chemicals *in vitro*. Environ Mol Mutagen 14:165–187 [cited in IARC 1999].

Izutani K, Nakata A, Shinagawa H, Kawamata J. 1980. Forward mutation assay for screening carcinogens by alkaline phosphatase constitutive mutations in *Escherichia coli* K-12. Biken J 23:69–75 [cited in IPCS 1996; IARC 1999].

Jacobs RS. 1985. Ethylene dibromide poisoning. J Am Med Assoc 253:2961 [cited in IPCS 1996].

Josephy PD, Taylor PL, Vervaet G, Mannervik B. 2006. Screening and characterization of variant theta-class glutathione transferases catalyzing the activation of ethylene dibromide to a mutagen. Environ Mol Mutagen 47:657–665.

Kale PG, Baum JW. 1979a. Sensitivity of *Drosophila melanogaster* to low concentrations of the gaseous 1,2-dibromoethane: I. Acute exposures. Environ Mutagen 1:15–18 [cited in IPCS 1996; IARC 1999].

Kale PG, Baum JW. 1979b. Sensitivity of *Drosophila melanogaster* to low concentrations of gaseous mutagens: II. Chronic exposures. Mutat Res 68:59–68 [cited in IPCS 1996; IARC 1999].

Kale PG, Baum JW. 1981. Sensitivity of *Drosophila melanogaster* to low concentrations of gaseous mutagens: III. Dose-rate effects. Environ Mutagen 3:65–70 [cited in IPCS 1996; IARC 1999].

Kale PG, Baum JW. 1982. Genetic effects of ethylene dibromide in *Drosophila melanogaster*. In: Tice RR, Costa DL, Schaich KM, editors. Genotoxic effects of airborne agents. New York (NY): Plenum Press. p. 291–300 [cited in IPCS 1996].

Kale PG, Baum JW. 1983. Sensitivity of *Drosophila melanogaster* to low concentrations of gaseous mutagens: IV. Mutations in embryonic spermatogonia. Mutat Res 113:135–143 [cited in IPCS 1996; IARC 1999].

Kale P, Kale R. 1995. Induction of delayed mutations by benzene and ethylene dibromide in *Drosophila*. Environ Mol Mutagen 25:211–215 [cited in IPCS 1996; IARC 1999].

Kerklaan P, Bouter S, Mohn G. 1983. Isolation of a mutant of *Salmonella typhimurium* strain TA1535 with decreased levels of glutathione (GSH–): primary characterization and chemical mutagenesis studies. Mutat Res 122:257–266 [cited in IPCS 1996].

Kerklaan P, Zoetemelk CEM, Mohn GR. 1985. Mutagenic activity of various chemicals in *Salmonella* strain TA100 and glutathione-deficient derivatives: on the role of glutathione in the detoxification or activation of mutagens inside bacterial cells. Biochem Pharmacol 34:2151–2156 [cited in IPCS 1996].

Kim DH, Guengerich FP. 1990. Formation of the DNA adduct *S*-[2-(N7-guanyl)ethyl] glutathione as an adduct formed in RNA and DNA from 1,2-dibromoethane. Chem Res Toxicol 3:587–594 [cited in IPCS 1996; IARC 1999].

Kitchin KT, Brown JL. 1994. Dose–response relationship for rat liver DNA damage caused by 49 rodent carcinogens. Toxicology 88:31–49 [cited in IARC 1999].

Klimisch HJ, Andreae M, Tillmann U. 1997. A systematic approach for evaluating the quality of experimental toxicological and ecotoxicological data. Regulatory Toxicology and Pharmacology. 25:1-5

Konishi Y, Yoshida S, Nakamura A. 1986. Determination of ethylene dibromide in foods and grains by high resolution capillary gas chromatography with electron capture detection. J Assoc Off Anal Chem 69(1):97–100.

Koptagel E, Bulut HE. 1998. Effects of short-term hydrocarbon inhalation on rat tracheal mucosa. Okajimas Folia Anat Jpn 75:71–86.

Krishna G, Xu J, Nath J, Petersen M, Ong T. 1985. *In vivo* cytogenetic studies on mice exposed to ethylene dibromide. Mutat Res 158:81–87 [cited in IPCS 1996; IARC 1999].

Kszos LA, Talmage SS, Morris GW, Konetsky BK, Rottero T. 2003. Derivation of aquatic screening benchmarks for 1,2-dibromoethane. Arch Environ Toxicol 45:66–71.

Kubinski H, Gutzke GE, Kubinski ZO. 1981. DNA-cell-binding (DCB) assay for suspected carcinogens and mutagens. Mutat Res 89:95–136 [cited in IPCS 1996].

Laturnus F. 1996. Volatile halocarbons released from Arctic macroalgae. Mar Chem 55(3–4):359–366.

Letz GA, Pond SM, Osterloh JD, Wade RL, Becker CE. 1984. Two fatalities after acute occupational exposure to ethylene dibromide. J Am Med Assoc 252(17):2428–2431 [cited in IPCS 1996; IARC 1999].

Marks CF, Thomason IJ, Castro CE. 1968. Dynamics of the permeation of nematodes by water, nematocides and other substances. Exp Parasitol 22:321–337.

McCann J, Choi E, Yamasaki E, Ames BN. 1975. Detection of carcinogens as mutagens in the *Salmonella*/microsome test: assay of 300 chemicals. Proc Natl Acad Sci U S A 72:5135–5139 [cited in IPCS 1996; IARC 1999].

McKay G. 1986. GC determination of EDB in flour products. J Assoc Off Anal Chem 68(2):203–205.

Mehrotra P, Naik SR, Choudhuri G. 2001. Two cases of ethylene dibromide poisoning. Vet Hum Toxicol 43:91–92.

Meneghini R. 1974. Repair replication of opossum lymphocyte DNA: effect of compounds that bind to DNA. Chem Biol Interact 8:113–126 [cited in IPCS 1996].

Mohn GR, Kerklaan PRM, Van Zeeland AA, Ellenberger J, Baan RA, Lohman PHM, Pons F-W. 1984. Methodologies for the determination of various genetic effects in permeable strains of *E. coli* K-12 differing in DNA repair capacity: quantification of DNA adduct formation, experiments with organ homogenates and hepatocytes, and animal-mediated assays. Mutat Res 125:153–184 [cited in IPCS 1996; IARC 1999].
Moriya M, Ohta T, Watanabe K, Miyazawa T, Kato K, Shirasu Y. 1983. Further mutagenicity studies on pesticides in bacterial reversion assay systems. Mutat Res 116:185–216 [cited in IPCS 1996; IARC 1999].

Nachtomi E, Sarma DSR. 1977. Repair of rat liver DNA *in vivo* damaged by ethylene dibromide. Biochem Pharmacol 26:1941–1945 [cited in IPCS 1996; IARC 1999].

Nakamura Y, Hasegawa Y, Tonogai Y, Hanafusa M, Hirose H, Taharasako Y, Ito Y. 1989. Analysis of EDB residue in the imported fruits. J Jpn Soc Food Sci Technol 36(2):142–147.

[NAPS] National Air Pollution Surveillance Network [database on the Internet]. 2008. Gatineau (QC): Environment Canada. [cited 2009 Aug. 4]. Available from: <u>http://www.etc-cte.ec.gc.ca/NAPS/index_e.html</u>

National Chemical Inventories [database on a CD-ROM]. 2006. Columbus (OH): American Chemical Society, Chemical Abstracts Service. Available from: http://www.cas.org/products/cd/nci/require.html

[NCI] National Cancer Institute. 1978. Bioassay of 1,2-dibromoethane for possible carcinogenicity. Bethesda (MD): National Institutes of Health, National Cancer Institute, Division of Cancer Cause and Prevention, Carcinogenesis Testing Program. 64 p. Technical Report Series No. 86. DHEW Publication No. (NIH) 78-1336.

Nitschke KD, Kociba RJ, Keyes DG, McKenna MJ. 1981. A thirteen week repeated inhalation study of ethylene dibromide in rats. Fundam Appl Toxicol 1:437–442 [cited in IPCS 1996; IARC 1999].

Novotná B, Duverger-van Bogaert M. 1994. Role of kidney S9 in the mutagenic properties of 1,2-dibromoethane. Toxicol Lett 74:255–263 [cited in IARC 1999].

[NPI] National Pollutant Inventory (AU). 2006. 1,2-Dibromoethane fact sheet. Canberra (AU): Australian Government, Department of the Environment, Water, Heritage and the Arts. Available from: <u>http://www.npi.gov.au/substances/dibromoethane/index.html</u>

[NPRI] National Pollutant Release Inventory [database on the Internet]. 2008. Gatineau (QC): Environment Canada. [cited 2009 Aug. 4]. Available from: http://www.ec.gc.ca/pdb/querysite/query_e.cfm

[NSEL] Nova Scotia Environment and Labour. 2005. 1,2-Dibromoethane (EDB) test results (mg/L). Sydney (NS): Nova Scotia Environment and Labour.

[NTP] National Toxicology Program (US). 1982. Carcinogenesis bioassay of 1,2-dibromoethane in F344 rats and B6C3F1 mice (inhalation study). Research Triangle Park (NC): US Department of Health and Human Services, National Toxicology Program. 163 p. NTP Technical Report Series No. 210.

[NTP] National Toxicology Program (US). 1992. NTP database search application: Study No. 738945. Research Triangle Park (NC): US Department of Health and Human Services, National Toxicology Program. Available from: <u>http://ntp-apps.niehs.nih.gov/ntp_tox/index.cfm</u>

[NTP] National Toxicology Program (US). 1993. NTP database search application: Study No. 738945. Research Triangle Park (NC): US Department of Health and Human Services, National Toxicology Program. Available from: <u>http://ntp-apps.niehs.nih.gov/ntp_tox/index.cfm</u>

[NTP] National Toxicology Program (US). 2005. 11th Report on carcinogens. Substance profile: 1,2-Dibromoethane (Ethylene Dibromide). Research Triangle Park (NC): National Toxicology Program. Available from: <u>http://ntp.niehs.nih.gov/ntp/roc/eleventh/profiles/s059dibr.pdf</u>

Oda Y, Yamazaki H, Thier R, Ketterer B, Guengerich FP, Shimada T. 1996. A new *Salmonella typhimurium* NM5004 strain expressing rat glutathione *S*-transferase 5-5: use in detection of genotoxicity of dihaloalkanes using an SOS/umu test system. Carcinogenesis 17:297–302 [cited in IARC 1999].

[OECD] Organisation for Economic Co-operation and Development. 2009. Manual for the Assessment of Chemicals. Annex 1: Guidance for Completing a SIDS Dossier. Paris (FR): OECD, Environment Directorate.

Ohta T, Nakamura N, Moriya M, Shirasu Y, Kada T. 1984. The SOS-function-inducing activity of chemical mutagens in *Escherichia coli*. Mutat Res 131:101–109 [cited in IPCS 1996].

[OMEE] Ontario Ministry of Environment and Energy. 1993. Ontario typical range of chemical parameters in soil, vegetation, moss bags and snow. Toronto (ON): Ontario Ministry of Environment and Energy, Standards Development Branch, Phytotoxicology Section. [cited 2009 Dec. 10]. Available from:

http://www.ene.gov.on.ca/envision/sudbury/ontario_typical_range/index.htm

[OMEE] Ontario Ministry of Environment and Energy. 1994. Windsor air quality study: ambient air monitoring activities. Toronto (ON): Ontario Ministry of Environment and Energy.

Ong T-M, Stewart JD, Wen Y-F, Whong W-Z. 1987. Application of SOS umu-test for the detection of genotoxic volatile chemicals and air pollutants. Environ Mutagen 9:171–176 [cited in IPCS 1996; IARC 1999].

Ong T-M, Stewart JD, Tucker JD, Whong W-Z. 1989. Development of an *in situ* test system for detection of mutagens in the workplace. In: Waters MD, Sandhu SS, Lewtas J, Claxton L, Strauss G, Nesnow S, editors. Short-term bioassays in the analysis of complex

environmental mixtures. IV. New York (NY): Plenum Publishing. p. 25–36 [cited in IPCS 1996].

[Ontario MOE] Ontario Ministry of the Environment. 1988. Drinking water surveillance program: Ottawa (Lemieux Island) water treatment plant. Annual report, 1987. Toronto (ON): Ontario Ministry of the Environment.

[Ontario MOE] Ontario Ministry of the Environment. 2006. Summary of ethylene dibromide data for raw water, treated water and distribution water sample locations as monitored by the Drinking Water Surveillance Program (DWSP), monitoring period 2005 to 2006. Toronto (ON): Ontario Ministry of the Environment, Environmental Monitoring and Reporting Branch, Water Monitoring and Reporting Section, Drinking Water Unit.

Otson R. 1986. Surveys of selected organics in residential air. In: Walkinshaw DS, editor. Indoor air quality in cold climates. Ottawa (ON): Air Pollution Control Association.

Ott MG, Scharnweber HC, Langner RR. 1980. Mortality experience of 161 employees exposed to ethylene dibromide in two production units. Br J Ind Med 37:163–168 [cited in IPCS 1996; IARC 1999].

Packer K. 1980. Nanogen index. Freedom (CA): Nanogens International. p. 47 [cited in Sawyer and Walters 1986].

Page GW. 1981. Comparison of groundwater and surface water for patterns and levels of contamination by toxic substances. Environ Sci Technol 15(12):1475–1481.

Patriarche J, Campbell ID. 1999. Alkyl lead inventory study. Sources, uses and releases in Ontario, Canada—a preliminary review. Prepared by Patriarche & Associates for Environmental Contaminants & Nuclear Programs Division, Environmental Protection Branch – Ontario Region, Environment Canada. 47 p.

[PCKOCWIN] Organic Carbon Partition Coefficient Program for Windows [Estimation Model]. 2008. Version 2.00. Washington (DC): US Environmental Protection Agency, Office of Pollution Prevention and Toxics; Syracuse (NY): Syracuse Research Corporation. Available from: <u>http://www.epa.gov/oppt/exposure/pubs/episuite.htm</u>

Peoples SA, Maddy KT, Riddle LC. 1978. Human occupational health problems resulting from exposure to ethylene dibromide in California in 1975 and 1976. Vet Hum Toxicol 20:241–244 [cited in IPCS 1996; IARC 1999].

Perocco P, Prodi G. 1981. DNA damage by haloalkanes in human lymphocytes cultured *in vitro*. Cancer Lett 13:213–218 [cited in IPCS 1996].

Perocco P, Colacci A, Santucci MA, Vaccari M, Grilli S. 1991. Transforming activity of ethylene dibromide in BALB/c 3T3 cells. Res Commun Chem Pathol Pharmacol 73(2):159–172 [cited in IPCS 1996; IARC 1999].

[PhysProp] Interactive PhysProp Database [database on the Internet]. 2009. Syracuse (NY): Syracuse Research Corporation. [cited 2009 Dec. 7]. Available from: http://esc.syrres.com/interkow/webprop.exe?CAS=106-93-4

Pignatello JJ. 1986. Ethylene dibromide mineralization in soils under aerobic conditions. Appl Environ Microbiol 51(3):588–592.

Pignatello JJ, Cohen SZ. 1990. Environmental chemistry of ethylene dibromide in soil and ground water. Rev Environ Contam Toxicol 112:1-47.

[PMRA] Pest Management Regulatory Agency. 2007. Regulatory Note REG 2007-04: PMRA list of formulants [Internet]. Ottawa (ON): Health Canada, Pest Management Regulatory Agency. [updated 2007 Jun; cited 2009 Aug 6]. Available from: http://www.hc-sc.gc.ca/cps-spc/pubs/pest/_decisions/reg2007-04/index-eng.php

[PPDB] Pesticide Properties Database [database on the Internet]. 2009. 1,2-Dibromoethane. European Commission. [updated 2009 Jul 27; cited 2009 Aug 4]. Available from: <u>http://sitem.herts.ac.uk/aeru/footprint/en/index.htm</u>

Prakash MS, Sud K, Kohli HS, Jha V, Gupta KL, Sakhuja V. 1999. Ethylene dibromide poisoning with acute renal failure: first reported case with non-fatal outcome. Ren Fail 21:219–222.

Principe P, Dogliotti E, Bignami M, Crebelli R, Falcone E, Fabrizi M, Conti G, Comba P. 1981. Mutagenicity of chemicals of industrial and agricultural relevance in *Salmonella*, *Streptomyces* and *Aspergillus*. J Sci Food Agric 32:826–832 [cited in IPCS 1996; IARC 1999].

Prodi G, Arfellini G, Colacci A, Grilli S, Mazzullo M. 1986. Interaction of halocompounds with nucleic acids. Toxicol Pathol 14(4):438–444 [cited in IARC 1999].

Qiu LX, Shi SH, Xing SB, Chen SG. 1992. Rate constants for the reactions of OH with five halogenated substituted ethanes from 292 K to 366 K. J Phys Chem 96:685–689.

Quillardet P, De Bellecombe C, Hofnung M. 1985. The SOS chromotest, a colorimetric bacterial assay for genotoxins: validation study with 83 compounds. Mutat Res 147:79–95 [cited in IPCS 1996; IARC 1999].

Rains D, Holder J. 1981. Ethylene dibromide residues in biscuits and flour. J Assoc Off Anal Chem 64(5):1252–1254.

Raman PG, Sain T. 1999. Clinical profile of ethylene di-bromide (EDB; 1,2-dibromo ethane) poisoning. J Assoc Physicians India 47:712–713.

Rannug U, Beije B. 1979. The mutagenic effect of 1,2-dichloroethane on *Salmonella typhimurium*. II. Activation by the isolated perfused rat liver. Chem Biol Interact 24:265–285 [cited in IPCS 1996].

Ratajczak HV, Aranyi C, Bradof JN, Barbera P, Fugmann R, Fenters JD, Thomas PT. 1994. Ethylene dibromide: evidence of systemic and immunologic toxicity without impairment of *in vivo* host defenses. In Vivo 8:879–884.

Ratajczak HV, Thomas PT, Gerhart J, Sothern RB. 1995. Immunotoxicologic effects of ethylene dibromide in the mouse and their modulation by the estrous cycle. In Vivo 9:299–304.

Ratcliffe JM, Schrader SM, Steenland K, Clapp DE, Turner T, Hornung RW. 1987. Semen quality in papaya workers with long term exposure to ethylene dibromide. Br J Ind Med 44:317–326 [cited in IPCS 1996; IARC 1999].

Reznik G, Stinson SF, Ward GM. 1980. Respiratory pathology in rats and mice after inhalation of 1,2-dibromo-3-chloropropane or 1,2-dibromoethane for 13 weeks. Arch Toxicol 46:233–240.

Roldán-Arjona T, Garcia-Pedrajas MD, Luque-Romero FL, Hera C, Pueyo C. 1991. An association between mutagenicity of the Ara test of *Salmonella typhimurium* and carcinogenicity in rodents for 16 halogenated aliphatic hydrocarbons. Mutagenesis 6:199–205 [cited in IARC 1999].

Rosenkranz HS. 1977. Mutagenicity of halogenated alkanes and their derivatives. Environ Health Perspect 21:79–84 [cited in IPCS 1996].

Rowe VK, Spencer HC, McCollister DD, Hollingsworth RL, Adams EM. 1952. Toxicity of ethylene dibromide determined on experimental animals. A M A Arch Ind Hyg Occup Med 6:158–173 [cited in IPCS 1996].

Russell WL. 1986. Positive genetic hazard predictions from short-term tests have proved false for results in mammalian spermatogonia with all environmental chemicals so far tested. In: Genetic toxicology of environmental chemicals—Part B: Genetic effects and applied mutagenesis. New York (NY): Alan R. Liss. p. 67–74 [cited in IPCS 1996].

Sarawat PK, Kandara M, Dhurva AK, Malhotra VK, Jhanwar RS. 1986. Poisoning by ethylene dibromide—six cases: a clinicopathological and toxicological study. Indian J Med Sci 40:121–123 [cited in IPCS 1996].

Sasaki YF, Saga A, Akasaka M, Ishibashi S, Yoshida K, Su YQ, Matsusaka N, Tsuda S. 1998. Detection of *in vivo* genotoxicity of haloalkanes and haloalkenes carcinogenic to rodents by the alkaline single cell gel electrophoresis (comet) assay in multiple mouse organs. Mutat Res 419:13–20.

Sawyer L, Walters S. 1986. Gas chromatographic method for ethylene dibromide in grains and grain-based products: collaborative study. J Assoc Off Anal Chem 69:847–851.

Schrader SM, Ratcliffe JM, Turner TW, Hornung RW. 1987. The use of new field methods of semen analysis in the study of occupational hazards to reproduction: the example of ethylene dibromide. J Occup Med 29:963–966 [cited in IPCS 1996].

Schrader SM, Turner TW, Ratcliffe JM. 1988. The effects of ethylene dibromide on semen quality: a comparison of short-term and chronic exposure. Reprod Toxicol 2:191–198 [cited in IPCS 1996; IARC 1999].

Scott BR, Sparrow AH, Schwemmer SS, Schairer LA. 1978. Plant metabolic activation of 1,2-dibromoethane (EDB) to a mutagen of greater potency. Mutat Res 49:203–212 [cited in IPCS 1996; IARC 1999].

SCREEN3 [Computer Model]. 1995. Version 96043.Research Triangle Park (NC): US Environmental Protection Agency, Office of Air Quality Planning and Standards, Emissions, Monitoring, and Analysis Division. Available from: <u>http://www.epa.gov/scram001/dispersion_screening.htm</u>

Sega GA, Sotomayor RE. 1980. Chemical dosimetry and unscheduled DNA synthesis studies of ethylene dibromide in the germ cells of male mice. Environ Mutagen 2:274 [incorrectly cited as Sega and Rene 1980 in IPCS 1996].

Shiau SY, Huff RA, Wells BC, Felkner IC. 1980. Mutagenicity and DNA-damaging activity for several pesticides tested with *Bacillus subtilis* mutants. Mutat Res 71:169–179 [cited in IPCS 1996].

Shivanandappa T, Krishnakumari MK, Majumder SK. 1987. Reproductive potential of male rats fed dietary ethylene dibromide. J Food Saf 8:147–155 [cited in IPCS 1996].

Short RD, Minor JL, Winston JM, Seifter J, Lee CC. 1978. Inhalation of ethylene dibromide during gestation by rats and mice. Toxicol Appl Pharmacol 46:173–182.

Short RD, Winston JM, Hong CB, Minor JL, Lee CC, Seifter J. 1979. Effects of ethylene dibromide on reproduction in male and female rats. Toxicol Appl Pharmacol 49:97–105 [cited in IPCS 1996; IARC 1999].

Simula TP, Glancey MJ, Wolf CR. 1993. Human glutathione *S*-transferase-expressing *Salmonella typhimurium* tester strains to study the activation/detoxification of mutagenic compounds: studies with halogenated compounds, aromatic amines and aflatoxin B1. Carcinogenesis 14:1371–1376 [cited in IPCS 1996; IARC 1999].

Sina JF, Bean CL, Dysart GR, Taylor VI, Bradley MO. 1983. Evaluation of the alkaline elution/rat hepatocyte assay as a predictor of carcinogenic/mutagenic potential. Mutat Res 113:357–391 [cited in IPCS 1996; IARC 1999].

Singh HB, Salas LJ, Stiles RE. 1982. Distribution of selected gaseous organic mutagens and suspect carcinogens in ambient air. Environ Sci Technol 16(12):872–880.

Singh N, Jatva OP, Gupta RK, Tailor MK, Jain R. 2007. Outcome of sixty-four cases of ethylene dibromide ingestion treated in tertiary care hospital. J Assoc Physicians India. 55:842-845.

Singh S, Chaudhry D, Garg M, Sharma BK. 1993. Fatal ethylene dibromide ingestion. J Assoc Physicians India 41:608 [cited in IARC 1999].

Smith RF, Goldman L. 1983. Behavioral effects of prenatal exposure to ethylene dibromide. Neurobehav Toxicol Teratol 5:579–585.

Steenland K, Carrano A, Ratcliffe J, Clapp D, Ashworth L, Meinhardt T. 1985. Cytogenetic studies in humans after short-term exposure to ethylene dibromide. J Occup Med 27:729–732 [cited in IARC 1999].

Steenland K, Carrano A, Ratcliffe J, Clapp D, Ashworth L, Meinhardt T. 1986. A cytogenetic study of papaya workers exposed to ethylene dibromide. Mutat Res 170:151–160 [cited in IPCS 1996; IARC 1999].

Stinson SF, Reznik G, Ward JM. 1981. Characteristics of proliferative lesions in the nasal cavities of mice following chronic inhalation of 1,2-dibromoethane. Cancer Lett 12:121–129 [cited in IPCS 1996; IARC 1999].

Stolzenberg SJ, Hine CH. 1980. Mutagenicity of 2- and 3-carbon halogenated compounds in the *Salmonella*/mammalian-microsome test. Environ Mutagen 2:59–66 [cited in IPCS 1996; IARC 1999].

Storer RD, Conolly RB. 1983. Comparative *in vivo* genotoxicity and acute hepatotoxicity of three 1,2-dihaloethanes. Carcinogenesis 4(11):1491–1494 [cited in IPCS 1996; IARC 1999].

Sugiyama H. 1980. Effects of EDB (1,2-dibromoethane) on the silkworm (*Bombyx mori* L.).J Pestic Sci 5:599–602 [cited in IPCS 1996].

Tan E-L, Hsie AW. 1981. Mutagenicity and cytotoxicity of haloethanes as studied in the CHO/HGPRT system. Mutat Res 90:183–191 [cited in IPCS 1996; IARC 1999].

Tananaki C, Zotou A, Thrasyvoulou A. 2005. Determination of 1,2-dibromoethane, 1,4-dichlorobenzene and naphthalene residues in honey by gas chromatography–mass

spectrometry using purge and trap thermal desorption extraction. J Chromatogr A 1083:146–152.

Tananaki C, Thrasyvoulou A, Karazafiris E, Zotou A. 2006. Contamination of honey by chemicals applied to protect honeybee combs from wax-moth (*Galleria mellonela* L.). Food Addit Contam 23(2):159–163.

[TaPL3] Long Range Transport and Persistence Level III model [Internet]. 2000. Version 2.10. Peterborough (ON): Trent University, Canadian Environmental Modelling Centre. Available from: <u>http://www.trentu.ca/academic/aminss/envmodel/models/TaPL3.html</u>

Teaf CM, Bishop JB, Harbison RD. 1990. Potentiation of ethyl methanesulfonateinduced germ cell mutagenesis and depression of glutathione in male reproductive tissues by 1,2-dibromoethane. Teratog Carcinog Mutagen 10:427–438 [cited in IARC 1999].

Tennant RW, Stasiewicz S, Spalding JW. 1986. Comparison of multiple parameters of rodent carcinogenicity and *in vitro* genetic toxicity. Environ Mutagen 8:205–227 [cited in IPCS 1996].

Tennant RW, Margolin BH, Shelby MD, Zeiger E, Haseman JK, Spalding J, Caspary W, Resnick M, Stasiewicz S, Anderson B, et al. 1987. Prediction of chemical carcinogenicity in rodents from *in vitro* genetic toxicity assays. Science 236:933–941 [cited in IPCS 1996].

Teramoto S, Saito R, Aoyama H, Shirasu Y. 1980. Dominant lethal mutation induced in male rats by 1,2-dibromo-3-chloropropane (DBCP). Mutat Res 77:71–78 [cited in IPCS 1996; IARC 1999].

Ter Haar G. 1980. An investigation of possible sterility and health effects from exposure to ethylene dibromide. In: Ames B, Infante O, Peirtz R, editors. Ethylene dichloride: a potential health risk? Cold Spring Harbor (NY): Cold Spring Harbor Laboratory. p. 167–188. Banbury Report No. 5 [cited in IPCS 1996].

Tezuka H, Ando N, Suzuki R, Terahata M, Moriya M, Shirasu Y. 1980. Sister-chromatid exchanges and chromosomal aberrations in cultured Chinese hamster cells treated with pesticides positive in microbial reversion assays. Mutat Res 78:177–191 [cited in IPCS 1996; IARC 1999].

Thier R, Pemble SE, Kramer H, Taylor JB, Guengerich FP, Ketterer B. 1996. Human glutathione *S*-transferase T1-1 enhances mutagenicity of 1,2-dibromoethane, bromomethane and 1,2,3,4-diepoxybutane in *Salmonella typhimurium*. Carcinogenesis 17:163–166 [cited in IARC 1999].

[TRI] Toxics Release Inventory [database on the Internet]. 2007. TRI Explorer 4.8. Washington (DC): US Environmental Protection Agency. [cited 2009 Aug. 4]. Available from: <u>http://www.epa.gov/triexplorer/</u> Tucker JD, Xu J, Stewart J, Ong T-M. 1984. Detection of sister-chromatid exchanges in human peripheral lymphocytes induced by ethylene dibromide vapor. Mutat Res 138:93–98 [cited in IPCS 1996; IARC 1999].

[UNEP and FAO] United Nations Environment Programme and Food and Agriculture Organization of the United Nations. 2003. PIC Circular XVIII – December 2003. Interim Secretariat for the Rotterdam Convention on the Prior Informed Consent Procedure for Certain Hazardous Chemicals and Pesticides in International Trade. Available from: <u>http://www.pic.int/en/Circular/CIRC18EN.pdf</u>

[US EPA] US Environmental Protection Agency. 1999. 1990 Emissions Inventory of Forty Potential Section 112(k) Pollutants. Research Triangle Park (NC): US Environmental Protection Agency, Emission Factors and Inventory Group (MD-14) and Emissions Standards Division (MD-15).

[US EPA] US Environmental Protection Agency. 2004. Toxicological review of 1,2dibromoethane (106-93-4) in support of summary information on the Integrated Risk Information System (IRIS). Available from: <u>http://www.epa.gov/ncea/iris</u>

[US FDA] US Food and Drug Administration. 2003. Food and Drug Administration total diet study: summary of residues found ordered by pesticide. Market baskets 91-3-01-4. College Park (MD): US Food and Drug Administration, Center for Food Safety and Applied Nutrition.

Van Bladeren PJ, Breimer DD, Rotteveel-Smijs GMT, De Jong RAW, Buus W, Van Der Gen A, Mohn GR. 1980. The role of glutathione conjugation in the mutagenicity of 1,2-dibromoethane. Biochem Pharmacol 29:2975–2982 [cited in IPCS 1996].

Van Bladeren PJ, Breimer DD, Rotteveel-Smijs GMT, De Knijff P, Mohn GT, Van Meeteren-Walchli B, Buijs W, Van der Gen A. 1981. The relation between the structure of vicinal dihalogen compounds and their mutagenic activation via conjugation to glutathione. Carcinogenesis 2(6):499–505 [cited in IPCS 1996; IARC 1999].

Van Duuren BL, Goldschmidt BM, Loewengart G, Smith AC, Melchlonne S, Seldman I, Roth D. 1979. Carcinogenicity of halogenated olefinic and aliphatic hydrocarbons in mice. J Natl Cancer Inst 63(6):1433–1439.

Van Duuren BL, Seidman I, Melchionne S, Kline SA. 1985. Carcinogenicity bioassays of bromoacetaldehyde and bromoethanol—potential metabolites of dibromoethane. Teratog Carcinog Mutagen 5:393–403.

Ville de Montréal. 2006. Municipal drinking water produced by Atwater and Charles-J Des Baillets drinking water plants. Montréal (QC): Division de l'expertise technique, Service des infrastructures et de l'environnement.

Vogel E, Chandler JLR. 1974. Mutagenicity testing of cyclamate and some pesticides in *Drosophila melanogaster*. Experientia 30:621–623 [cited in IPCS 1996; IARC 1999].

Vogel TM, Reinhard M. 1986. Reaction products and rates of disappearance of simple bromoalkanes, 1,2-dibromopropane and 1,2-dibromoethane in water. Environ Toxicol Chem 7:917–924.

Von Buselmaier W, Rohrborn G, Propping P. 1972. [Mutagenicity investigations with pesticides in the host mediated assay and the dominant lethal test in mice.] Biol Zent Bl 91:311–325 [in German] [cited in IPCS 1996].

Walton BT, Hendricks MS, Anderson TA, Talmage SS. 1989. Treatability of hazardous chemicals in soils: volatile and semivolatile organics. Oak Ridge (TN): Environmental Sciences Division, Oak Ridge National Laboratory. (Publication No. 3283).

Watanabe K, Sasaki T, Kawakami K. 1998. Comparisons of chemically-induced mutation among four bacterial strains, *Salmonella typhimurium* TA102 and TA2638, and *Escherichia coli* WP2/pKM101 and WP2 *uvrA*/pKM101: collaborative study III and evaluation of the usefulness of these strains. Mutat Res 416:169–181.

Watanabe K, Liberman RG, Skipper PL, Tannenbaum SR, Guengerich FP. 2007. Analysis of DNA adducts formed *in vivo* in rats and mice from 1,2-dibromoethane, 1,2-dichloroethane, dibromomethane, and dichloromethane using HPLC/accelerator mass spectrometry and relevance to risk estimates. Chem Res Toxicol 20:1594–1600.

Weintraub RA, Jex GW, Moye HA. 1986. Chemical and microbial degradation of 1,2-dibromoethane (EDB) in Florida ground water, soil and sludge. Washington (DC): American Chemical Society. p. 294–310.

White RD. 1982. Chemical induction of genetic injury: the bioactivation of 1,2-dibromoethane. Diss Abstr Int 43(03):696B–697B [cited in IPCS 1996].

White RD, Sipes IG, Gandolfi AJ, Bowden GT. 1981. Characterization of the hepatic DNA damage caused by 1,2-dibromoethane using the alkaline elution technique. Carcinogenesis 2:839–844 [cited in IARC 1999].

Wildeman AG, Nazar RN. 1982. Significance of plant metabolism in the mutagenicity and toxicity of pesticides. Can J Genet Cytol 24:437–449 [cited in IPCS 1996].

Williams GM, Laspia MF, Dunkel VC. 1982. Reliability of the hepatocyte primary culture/DNA repair test in testing of coded carcinogens and noncarcinogens. Mutat Res 97:359–370 [cited in IPCS 1996; IARC 1999].

Williams WM, Holden PW, Parsons DW. 1988. Pesticides in Groundwater Data Base: 1988 Interim Report. Washington (DC): US Environmental Protection Agency. NTIS no. PB89 164230.

Witt KL, Knapton A, Wehr CM, Hook GJ, Mirsalis J, Shelby MD, MacGregor JT. 2000. Micronucleated erythrocyte frequency in peripheral blood of B6C3F1 mice from shortterm, prechronic, and chronic studies of the NTP Carcinogenesis Bioassay Program. Environ Mol Mutagen 36:163–194.

Wong LCK, Winston JM, Hong CB, Plotnick H. 1982. Carcinogenicity and toxicity of 1,2-dibromoethane in the rat. Toxicol Appl Pharmacol 63:155–165.

Wong O, Morgan RW, Whorton MD. 1985. An epidemiologic surveillance program for evaluating occupational reproductive hazards. Am J Ind Med 7:295–306.

Working PK, Smith-Oliver T, White RD, Butterworth BE. 1986. Induction of DNA repair in rat spermatocytes and hepatocytes by 1,2-dibromoethane: the role of glutathione conjugation. Carcinogenesis 7(3):467–472 [cited in IPCS 1996; IARC 1999].

Yoshida YH, Inagaki E. 1986. [Mutagenicity of ethylene dibromide in *Drosophila melanogaster*.] Tachikawa Tandai Kiyo 19:49–50 [in Japanese] [cited in IPCS 1996].

Zeiger E, Anderson B, Haworth S, Lawlor T, Mortelman K. 1992. *Salmonella* mutagenicity tests V. Results from the testing of 311 chemicals. Environ Mol Mutagen 19 (Suppl 21):2–141.

Zhu J, Newhook R, Marro L, Chan CC. 2005. Selected volatile organic compounds in residential air in the City of Ottawa, Canada. Environ Sci Technol 39:3964–3971.

Zoetemelk CEM, Mohn GR, Van Der Gen A, Breimer DD. 1987. Mutagenicity in *Salmonella* strains differing in glutathione content and their alkylating potential. Biochem Pharmacol 36(11):1829–1835 [cited in IPCS 1996].

Zogorski JS, Carter JM, Ivahnenko T, Lapham WW, Moran MJ, Rowe BL, Squillace PJ, Toccalino PL. 2006. The quality of our nation's waters—Volatile organic compounds in the nation's ground water and drinking-water supply wells [Internet]. Reston (VA): US Department of the Interior, US Geological Survey. Circular 1292.

Appendix 1: Robust Study Summaries for Ecotoxicity Studies

Description of the reliability evaluation

For determination of the reliability of experimental data for key ecological endpoints (i.e., inherent toxicity to aquatic organisms, bioaccumulation potential, persistence), an evaluation approach has been developed, which is analogous to that of Klimisch et al. (1997). It involves the use of a standardized Robust Study Summary form, including a scoring system to quantitatively evaluate the studies.

The Robust Study Summary (RSS) is an adaptation of the OECD Robust Study Summary templates (OECD 2009). It consists of a checklist of items or criteria relating to identity of the substance, experimental protocol or method, test organism, specific test design/conditions, ecological relevance, and results. Most items are weighted according to their criticality to the quality and reliability of the study. The most important or critical items (which describe parameters/factors that have the most direct influence on the quality of the study) have been given a higher weight (5 points), while the less critical items have been given a lower score (1 or 2 points). For each item, the evaluator must indicate whether the item has been addressed appropriately in the study by answering "yes", "no" or "non-applicable (n/a)".Specific information relating to the items is also provided the RSS as well.

Once answers to all the items have been provided in the template, an overall Robust Study Summary score for the study is calculated as:

Overall Study Score (%) =
$$\frac{\sum W_{Yes}}{\sum W_{Yes+No}} \times 100\%$$

Where: W_{Yes} = weight of applicable "Yes" answers; W_{Yes+No} = weight of applicable "Yes" and "No" answers.

The overall score's corresponding reliability code and category is determined using the four categories adapted from the Klimisch approach and based on the score ranges as described in Table A.

Reliability Code	Reliability Category	Overall Study Score Range
1	High confidence	≥ 80%
2	Satisfactory confidence	60 - 79%
3	Low confidence	40 - 59%
4	Not acceptable	< 40%

Table A	Coordina	Cuid for	0	C4 J	Dallah lita
Table A.	Scoring	Gria lor	Overall	Sludy	Renadinty

Study 1

No	Item	Weight	Yes/No	Specify						
1	Reference: Holcombe GW, Benoit 1995. Acute and long-term effects	DA, Hammo of nine cher	ermeister D nicals on th	E, Leonard EN, Johnson RD. ne Japanese medaka (<i>Oryzias</i>						
2	Substance identity: CAS RN	n/a	Y							
3	Substance identity: 1,2-dibromoethane	n/a	Y							
4	Chemical composition of the substance	2	Y							
5	Chemical purity	1	Y							
6	Persistence/stability of test substance?	1	Y							
	Method									
7	Reference	1	Y							
8	OECD, EU, national, or other standard method?	3	-	Not applicable						
9	Justification of the method/protocol if a non- standard method was used	2	Y							
10	GLP (good laboratory practice)	3	-	Not applicable						
	<i>T</i>	est organis	sm							
11	Organism identity: medaka	n/a	Y							
12	Latin or both Latin and common names reported?	1	Ν							
13	Life cycle age / stage of test organism	1	Y	28–43 days old for acute tests						
14	Length and/or weight	1	Y	18–71 mg						
15	Sex	1	N							
16	Number of organisms per replicate	1	Y	10						
17	Organism loading rate	1	Y							
18	Food type and feeding periods during the acclimation period	1	Y							
	Test d	lesign / con	ditions							
19	Test type (acute or chronic)	n/a	Y	Both						
20	Experiment type (laboratory or field)	n/a	Y	Lab						
21	Exposure pathways (food, water, both)	n/a	Y	Water						
22	Exposure duration	n/a	Y	Acute 96h						
23	Negative or positive controls (specify)	1	Y							
24	Number of replicates (including controls)	1	Y							
25	Nominal concentrations reported?	1	Y							
26	Measured concentrations reported?	3	Y							
27	Food type and feeding periods during the long-term tests	1	Y							
28	Were concentrations measured periodically (especially in the chronic test)?	1	Y							
29	Were the exposure media conditions relevant to the particular chemical reported? (e.g., for the metal toxicity – pH,	3	Y							

	DOC/TOC, water hardness, temperature)			
30	Photoperiod and light intensity	1	Y	
31	Stock and test solution preparation	1	Y	
32	Was solubilizer/emulsifier used if the chemical was poorly soluble or unstable?	1	-	Not applicable
33	If solubilizer/emulsifier was used, was its concentration reported?	1	-	Not applicable
34	If solubilizer/emulsifier was used, was its ecotoxicity reported?	1	-	Not applicable
35	Monitoring intervals (including observations and water quality parameters) reported?	1	Ν	
36	Statistical methods used	1	Y	
	Information re	elevant to t	he data qu	ality
37	Was the endpoint directly caused by the chemical's toxicity, not by the organism's health (e.g. when mortality in the control > 10%) or physical effects (e.g. "shading effect")?	n/a	Y	
38	Was the test organism relevant to the Canadian environment?	3	Y	
39	Were the test conditions (pH, temperature, DO, etc.) typical for the test organism?	1	Y	
40	Do system type and design (static, semi-static, flow-through; sealed or open; etc.) correspond to the substance's properties and organism's nature/habits?	2	Y	
41	Was pH of the test water within the range typical for the Canadian environment (6 to 9)?	1	Y	
42	Was temperature of the test water within the range typical for the Canadian environment (5 to 27°C)?	1	Y	
43	Was toxicity value below the chemical's water solubility?	3	Y	
		Results		
44	Toxicity values (specify endpoint and value)	n/a	Y	
45	Other endpoints reported – e.g., BCF/BAF, LOEC/NOEC (specify)?	n/a	Ν	
46	Other adverse effects (e.g. carcinogenicity, mutagenicity) reported?	n/a	Ν	
47	Score: %			36/39 = 92%
48	EC reliability code:			1
49	Reliability category (high, satisfactory, low):			High
50	Comments			

Study 2

No	Item	Weight	Yes/No	Specify
1	Reference: Kszos LA, Talmage	SS, Morris	s GW, Kon	etsky BK, Rottero T. 2003. Derivation of
1	aquatic screening benchmarks	for 1,2-dib	romoethan	e. Arch Environ Toxicol 45:66-71.
2	Substance identity: CAS RN	n/a	Y	
3	Substance identity: 1,2- dibromoethane	n/a	Y	
4	Chemical composition of the substance	2	Y	
5	Chemical purity	1	-	Not specified but not needed
6	Persistence/stability of test substance?	1	Y	
		Met	hod	
7	Reference	1	Y	
8	OECD, EU, national, or other standard method?	3	-	Not applicable
9	Justification of the method/protocol if a non-standard method was used	2	Y	
10	GLP (good laboratory practice)	3	-	Not applicable
		Test or	ganism	
11	Organism identity: <i>Daphnia</i> <i>magna</i> , <i>Ceriodaphnia dubia</i> and <i>Pimephales promelas</i> (fathead minnow)	n/a	Y	
12	Latin or both Latin and common names reported?	1	Y	
13	Life cycle age / stage of test organism	1	Y	Fish (5 days old)
14	Length and/or weight	1	N	
15	Sex	1	N	
16	Number of organisms per replicate	1	Y	8
17	Organism loading rate	1	Y	5 concentrations on <i>D. magna</i> and <i>C. dubia</i> 4 concentrations on fish
18	Food type and feeding periods during the acclimation period	1	Y	
	Те	st design	/ conditio	ons
19	Test type (acute or chronic)	n/a	Y	Acute
20	Experiment type (laboratory or field)	n/a	Y	Lab
21	Exposure pathways (food, water, both)	n/a	Y	water
22	Exposure duration	n/a	Y	48 h for <i>D. magma</i> and <i>C. dubia</i> 98 h for fathead minnow
23	Negative or positive controls (specify)	1	Y	Negative control
24	Number of replicates (including controls)	1	Y	4 (for <i>D. magna</i> and <i>C. dubia</i>) 8 (fathead minnow)
25	Nominal concentrations reported?	1	Y	5 for <i>D. magna</i> and <i>C. dubia</i> 4 for P. promelas
26	Measured concentrations reported?	3	Y	

27	Food type and feeding periods during the long-term tests	1	Y	Feeding fish with brine shrimp nauplii 2 h prior to test solution renewal in 48 h
28	Were concentrations measured periodically (especially in the chronic test)?	1	Y	At least 2 times
29	Were the exposure media conditions relevant to the particular chemical reported? (e.g., for the metal toxicity – pH, DOC/TOC, water hardness, temperature)	3	Y	
30	Photoperiod and light intensity	1	Y	
31	Stock and test solution preparation	1	Y	
32	Was solubilizer/emulsifier used if the chemical was poorly soluble or unstable?	1	-	Not applicable
33	If solubilizer/emulsifier was used, was its concentration reported?	1	-	Not applicable
34	If solubilizer/emulsifier was used, was its ecotoxicity reported?	1	-	Not applicable
35	Monitoring intervals (including observations and water quality parameters) reported?	1	Y	
36	Statistical methods used	1	Y	
	Informatio	on relevan	t to the da	ata quality
37	Was the endpoint directly caused by the chemical's toxicity, not by organism's health (e.g. when mortality in the control > 10%) or physical effects (e.g. "shading effect")?	n/a	Y	
38	Was the test organism relevant to the Canadian environment?	3	Y	
39	Were the test conditions (pH, temperature, DO, etc.) typical for the test organism?	1	Y	
40	Do system type and design (static, semi-static, flow-through; sealed or open; etc.) correspond to the substance's properties and organism's nature/habits?	2	Y	
41	Was pH of the test water within the range typical for the Canadian environment (6 to 9)?	1	Y	
42	Was temperature of the test water within the range typical for the Canadian environment (5 to 27°C)?	1	Y	
43	Was toxicity value below the chemical's water solubility?	3	Y	
		Res	sults	
44	Toxicity values (specify	n/a	Y	

	endpoint and value)			
45	Other endpoints reported – e.g., BCF/BAF, LOEC/NOEC (specify)?	n/a	N	
46	Other adverse effects (e.g. carcinogenicity, mutagenicity) reported?	n/a	N	
47	Score: %			37/39 = 95%
48	EC Reliability code:			1
49	Reliability category (high, satisfactory, low):	High		
50	Comments			

Study 3

No	Item	Weight	Yes/No	Specify
1	Reference: Hawkins WE, Wa Overstreet RM. 1998. Carcinog EDB) in Jananese medaka (Or	ker WW, enic effect	James MO ts of 1,2-di	, Manning CS, Barnes DH, Heard CS, ibromoethane (ethylene dibromide; Res 399(2):221-32
2	Substance identity: CAS RN	n/a	N	(cs 399(2).221-32.
3	Substance identity: 1,2- dibromoethane	n/a	Y	
4	Chemical composition of the substance	2	Ν	Not specified but not needed
5	Chemical purity	1	Ν	Not specified but not needed
6	Persistence/stability of test substance?	1	Ν	
		Met	hod	
7	Reference	1	Y	
8	OECD, EU, national, or other standard method?	3	-	Not applicable
9	Justification of the method/protocol if a non-standard method was used	2	Y	
10	GLP (good laboratory practice)	3	-	Not applicable
		Test or	ganism	
11	Organism identity: Japanese medaka	n/a	Y	
12	Latin or both Latin and common names reported?	1	Ν	
13	Life cycle age / stage of test organism	1	Y	Fish (7 days old)
14	Length and/or weight	1	Ν	
15	Sex	1	N	
16	Number of organisms per replicate	1	Y	350
17	Organism loading rate	1	Y	1 static control 1 flow-through control 3 test concentrations
18	Food type and feeding periods during the acclimation period	1	Y	

	Test design / conditions					
19	Test type (acute or chronic)	n/a	Y	Chronic		
20	Experiment type (laboratory or field)	n/a	Y	Lab		
21	Exposure pathways (food, water, both)	n/a	Y	Water		
22	Exposure duration	n/a	Y	73-97 days		
23	Negative or positive controls (specify)	1	Y	Negative control		
24	Number of replicates (including controls)	1	Y	350		
25	Nominal concentrations reported?	1	Y	1 flow-through control 3 test concentrations		
26	Measured concentrations reported?	3	Y			
27	Food type and feeding periods during the long-term tests	1	Y	Very detailed		
28	Were concentrations measured periodically (especially in the chronic test)?	1	Y	Twice every week		
29	Were the exposure media conditions relevant to the particular chemical reported? (e.g., for the metal toxicity – pH, DOC/TOC, water hardness, temperature)	3	Y			
30	Photoperiod and light intensity	1	N			
31	Stock and test solution preparation	1	Y			
32	Was solubilizer/emulsifier used if the chemical was poorly soluble or unstable?	1	-	Not applicable		
33	If solubilizer/emulsifier was used, was its concentration reported?	1	-	Not applicable		
34	If solubilizer/emulsifier was used, was its ecotoxicity reported?	1	-	Not applicable		
35	Monitoring intervals (including observations and water quality parameters) reported?	1	Y			
36	Statistical methods used	1	Y			
	Informatio	on relevar	nt to the d	ata quality		
37	Was the endpoint directly caused by the chemical's toxicity, not by organism's health (e.g. when mortality in the control > 10%) or physical effects (e.g. "shading effect")?	n/a	Y			
38	Was the test organism relevant to the Canadian environment?	3	Y			
39	Were the test conditions (pH, temperature, DO, etc.) typical for the test organism?	1	Y			

40	Do system type and design (static, semi-static, flow-through; sealed or open; etc.) correspond to the substance's properties and organism's nature/habits?	2	Y	
41	Was pH of the test water within the range typical for the Canadian environment (6 to 9)?	1	Y	
42	Was temperature of the test water within the range typical for the Canadian environment (5 to 27°C)?	1	Y	
43	Was toxicity value below the chemical's water solubility?	3	Y	
		Res	sults	
44	Toxicity values (specify endpoint and value)	n/a	Y	
45	Other endpoints reported – e.g., BCF/BAF, LOEC/NOEC (specify)?	n/a	N	
46	Other adverse effects (e.g. carcinogenicity, mutagenicity) reported?	n/a	Y	
47	Score: %	35/40 = 87.5%		
48	EC Reliability code:			1
49	Reliability category (high, satisfactory, low):	High		
50	Comments			

Appendix 2: Concentrations of 1,2-dibromoethane in ambient air

Location	Sampling period	Number of samples	Detection limit (µg/m ³)	Mean concentration (µg/m ³) ¹	Reference
Halifax Nova	January to April, 2009	287	0.025	ND (ND-0.025)	Health Canada 2012
Scotia	June to September, 2009	324	0.025	ND (ND-0.026)	Health Canada 2012
Windsor,	January 23 to March 25, 2006	214	0.15	ND	Health Canada
Ontario	July 3 to August 26, 2006	214	0.15	ND	2010b
Windsor,	January 24 to March 19, 2005	201	0 123	ND	Health Canada
Ontario	July 4 to August 27, 2005	216	0.125	ND	2010b
Pagina	January 8 to March 16,	94(winter; 24-h		ND	Health Canada
Saskatchewan	Saskatchewan June 20 to August 29, 2007	97 (summer; 5-day canisters)	0.054	ND	2010a
43 Canadian sites	January to December 2008	10–119 (total of 1896 samples)	0.012	0.006 ³ (0.002– 0.013) [detected in 7 samples]	NAPS 2008
Twenty-nine Canadian cities	2004–2009	-	-	0-0.060	Environment Canada 2009b
Twenty-nine Canadian cities	1998–2002	-	-	< 0.012-0.143	Environment Canada 2004
40 Canadian sites	January to December 2003	14–145 (total of 1854 samples)	0.012	(ND–0.11) [detected in 458 samples]	2003 personal communication from Analysis and Air Quality Division, Environment Canada to Existing Substances Division; unreferenced
Ottawa, Ontario (vicinity of 75 homes)	Fall 2002	75	0.018	ND	Zhu et al. 2005

Location	Sampling	Number of	Detection limit	Mean concentration	Reference
	period	samples	$(\mu g/m^3)$	$(\mu g/m^3)^1$	
50 Canadian sites	January 1998 to December 2002	14–293 (total of 8275 samples)	0.012	(0.002–0.143) [detected in 6766 samples]	2002 personal communication from Analysis and Air Quality Division, Environment Canada to Existing Substances Division; unreferenced
37 Canadian sites	2000	9–62 (total of 1573 samples)	0.012	0.06 (0.01–0.12)	2001 personal communication from Analysis and Air Quality Division, Environment Canada to Existing Substances Division; unreferenced
Montréal, Quebec (urban)	1993	160	0.38 ² (0.05 ppbv)	0.02 (ND-0.67) [6% > detection limit]	Environment Canada 1995
Brossard, Quebec (suburban)	1993	24	0.38 ² (0.05 ppbv)	ND	Environment Canada 1995
Sainte- Françoise, Quebec (rural)	1993	34	0.38 ² (0.05 ppbv)	ND	Environment Canada 1995
Montréal, Quebec (urban)	1992	166	0.38 ² (0.05 ppbv)	0.01 (ND-1.73) [1% > detection limit]	Environment Canada 1995
Montréal, Quebec (urban)	1991	91	0.38 ² (0.05 ppbv)	0.03 (ND-0.48) [10% > detection limit]	Environment Canada 1995
Montréal, Quebec (urban)	1990	110	0.38 ² (0.05 ppbv)	ND-0.12 [1% > detection limit]	Environment Canada 1995
Montréal, Quebec (urban)	1989	79	0.38 ² (0.05 ppbv)	0.03 (ND-0.43) [11% > detection limit]	Environment Canada 1995

Location	Sampling period	Number of samples	Detection limit (µg/m ³)	Mean concentration (µg/m ³) ¹	Reference
Modelled local air dispersion (at 100 m from the source)	-	-	-	0.3774	SCREEN3 1995
Windsor, Ontario	1988–1992	410	0.1	ND-0.80 [ND 80% of the time]	OMEE 1994
Greater Vancouver Regional District	1989–1992	473	0.38 ² (0.05 ppbv)	0.06 ³ [4% > detection limit]	Environment Canada 1994
Canada-wide	1989–1990	1100	0.38^2 (0.05 ppbv)	0.06^3 [5% > detection limit]	Environment Canada 1994
Walpole Island, Ontario	1989–1991	94	0.1	ND-0.76	OMEE 1994
Walpole Island, Ontario	January 1988 to October 1990	61	0.1	ND–0.80 [above detection limit in 9 samples]	Environment Canada 1992
Windsor, Ontario	July 1987 to October 1990	123	0.1	ND–0.4 [above detection limit in 7 samples]	Environment Canada 1992
Canadian urban sites	1989	17	0.1	ND	Environment Canada 1991
Kitchener, Ontario	April 16 to May 24, 1989	10	ns	(ND-0.30)	CMHC 1989
North and South Atlantic Ocean	1985	0	0	0.020	Class and Ballschmiter 1988
Seven U.S. cities	1980	-	-	0.122-2.822	Singh et al. 1982

Abbreviations: ND, not detected; ns, not specified; ppbv, parts per billion by volume. ¹ Values in parentheses indicate range of concentrations when available. ² Value given for the detection limit is the target or typical detection limit reported for volatile organic compounds.
³ Values below the detection limit set at one-half the detection limit.

Appendix 3: Concentrations of 1,2-dibromoethane in indoor air

Location	Sampling period	Number of samples	Detection limit (μg/m ³)	Mean concentration (µg/m ³) ¹	Reference	
Halifax Nova Sootia	January to April, 2009	312	0.025	ND	Health Canada 2012	
namax, Nova Scolla	June to September, 2009	331	0.025	ND	Health Canada 2012	
Windsor, Ontario	January 24 to March 19, 2005	225	0 123	ND	Health	
(personal breathing-zone air)	July 4 to August 27, 2005	207	0.125	ND (ND-0.190)	2010b	
Windsor Ontario	January 24 to March 19, 2005	232	0 123	ND	Health	
windsor, Ontario	July 4 to August 27, 2005	217	0.125	ND	2010b	
Windsor, Ontario	January 23 to March 25, 2006	224	0 15	ND	Health	
	July 3 to August 26, 2006	211	015	ND	2010b	
Regina, Saskatchewan (5-day canister data were	January 8 to March 16, 2007	97(winter)		ND	Health	
selected, as they represent time-weighted average over longer period than 24-h canisters)	June 20 to August 29, 2007	101 (summer)	0.054	ND [maximum 0.080]	Canada 2010a	
Ottawa, Ontario (75 homes)	Fall 2002	75	0.018	ND	Zhu et al. 2005	
International locations (literature review of 50 studies)	1978–1990	50 studies	ns	1-<5	Brown et al. 1994	
Canada-wide	August– October and January– March 1983– 1984	10	0.4	ND	Otson 1986	

Location	Sampling period	Number of samples	Detection limit (µg/m ³)	Mean concentration (µg/m ³) ¹	Reference
Woodlands, California, USA (residential)	June 1990	128	ns	Not quantifiable	Cal EPA 1992
Kanawha Valley, West Virginia, USA (residential)	August 1987	35	8.5	6.06 [maximum 23.53; 29% > detection limit]	Cohen et al. 1989

Abbreviations: ND, not detected; ns, not specified. ¹ Values in parentheses indicate range of concentrations when available.

Appendix 4: Concentrations of 1,2-dibromoethane in water

Location	Sampling	pling Number of		Mean concentration	Reference
Location	period	samples	$(\mu g/L)$	(µg/L)	Reference
Victoria, British Columbia (drinking water)	2008	2	0.005	ND	City of Victoria 2008
Ontario, Canada (drinking water)	January 1, 2005 to December 31, 2006	2901	0.1	ND	Ontario MOE 2006
Montréal, Quebec (drinking water)	2006	ns	0.04	ND	Ville de Montreal 2006
Nova Scotia, Canada (drinking water)	June 2002 to May 2005	24	1	ND	NSEL 2005
Ottawa, Ontario (drinking water)	2003	19	0.10	ND	COWQS 2003
United States	1985–2001	462 (public well samples)	ns	< 0.10 (median for all samples)	Zogorski et al. 2006
United States	1985–2001	2085 (domestic well samples)	ns	< 0.04 (median for all samples); 0.55 (median for samples with detection)	Zogorski et al. 2006
United States	1985–2001	2851 (groundwater)	ns	< 0.10 (median for all samples); 0.72 (median for samples with detection)	Zogorski et al. 2006
Lemieux Island, Ottawa, Ontario	1987	48 (raw and treated)	50	Not quantifiable at detection limit	Ontario MOE 1988
Toronto, Ontario	November– December 1988	7 (bottled) 27 (tap)	0.04	Not quantifiable at detection limit	City of Toronto 1990
New Jersey, USA (surface water)	1977–1979	-	-	0.2 (maximum)	Page 1981
Oil refining and manufacturing facility, Sugar Creek, Missouri, USA (surface water)	1975 or earlier	-	-	1.05 – 1.13	Going and Long 1975
Pincher Creek, Alberta, at 50 m from the source (surface water)	-	-	-	16–21 ^a 2–3 ^b	ChemSim 2003

North and South Atlantic Ocean (marine water)	1985	-	-	0.00002	Class and Ballschmiter 1988
Site of a chemical plant, Ontario (groundwater)	1997	-	-	5.0	Environment Canada 2001b
Three U.S. states ^c (groundwater)	1981 – 1987	-	-	Detected	Pignatello and Cohen 1990
Six U.S. states ^d (groundwater)	1988 or earlier	-	-	14 (maximum) 9 (median)	Williams et al. 1988
New Jersey, USA (groundwater)	1977–1979	-	-	48 (maximum)	Page 1981

Abbreviations: ND, not detected; ns, not specified.

^a Most conservative scenarios: modelled value based on the assumption that the total amount reported in Canada is used at the Pincher Creek, Alberta, facility, with or without sewage treatment plant removal.

b More realistic scenarios (less conservative): modelled value based on the assumption that the total amount reported in Canada is divided among eight facilities, with or without sewage treatment plant removal.

^c Arizona, Wisconsin and Florida.
 ^d California, Connecticut, Georgia, Massachusetts, New York and Washington.

Appendix 5: Concentrations of 1,2-dibromoethane in food

Item sampled	Sampling period	Number of samples	Detection limit (µg/kg)	Mean concentration ¹ (µg/kg)	Reference	
Greece (domestic honey)	2004	25	0.8	Quantified in only two samples: 75 ± 3 and 12 ± 0.5	Tananaki et al. 2005	
	2003	142		Maximum 132.5		
Greece (honey)	2004	737		Maximum 331.2		
	2005	266		Maximum 95.2	Tananalsi at al	
fir honey		24	0.8	Maximum 12.7	2006	
blossom honey	2003 2005	60		Maximum 10.5	2000	
thymus honey	2003-2003	49		Maximum 2.9		
pine honey		283		Maximum 16.0		
United States (sweet cucumber pickles)	September– October 1991 to July– August 2001	1	0.5	13	US FDA 2003	
United States						
- cottage cheese				0.9 (ND-2.7)		
- popcorn in oil				0.4 (ND-1.3)		
- onion rings, breaded/fried				0.3 (ND-1.0)		
- frozen fried chicken				0.6 (ND-1.9)		
- honey, bottled				0.7 (ND-2.0)		
- chocolate cake/icing	April 1982 to April 1986	16	16 LOQ 1.0 ²	$LOQ 1.0^2$	4.0 (ND-11.9)	Gunderson
- yellow cake				2.8 (ND-8.4)	1900a	
- doughnuts				2.2 (ND-6.5)		
- cookies, chocolate chip			2.4 (ND-7.3)			
- cookies, sandwich				0.6 (ND-1.9)		
- apple pie, frozen				0.3 (ND-1.0)		
carbonated soda				0.003 (ND-		
				0.0084)		
				Pulp: 1.84 (ND–5.3)		
Florida (grapefruit)	April–June 1987	5	0.5	Peel: 3.12 (0.6–10.0)	Nakamura et al. 1989	
				Seeds: 336 (ND–591)		

Item sampled	Sampling period	Number of samples	Detection limit (µg/kg)	Mean concentration ¹ (µg/kg)	Reference
				Pulp: 0.95	
				(0.6–1.3)	
Israel (grapefruit)	April–June 1987	2	0.5	Peel: ND	Nakamura et al. 1989
				Seeds: 776 (521–1031)	
				Pulp: 1.57	
				(ND-2.8)	
Philippines (mango)	April–June 1987	6	0.5	Peel: 4.4 (2.7–6.3)	Nakamura et al. 1989
				Seeds: 2.47	
				(ND-4.1)	
				Pulp: 4.1	
				(ND-7.9)	
	Annil Juno			Deal: 6.4	Naltamura at
Mexico (mango)	Aprii–June 1987	4	0.5	$(ND_{15} 6)$	Nakamura et
	1987			(11D-13.0)	al. 1989
				Seeds: 58	
				(2.3–137)	
				Pulp: 0.75	
				(ND-2.4)	
				P 1 0 44	Nakamura et
Hawaii (papaya)	April–June	10	0.5	Peel: 0.66	al. 1989
	1987			(ND-1.5)	
				Seeds: 1.0	
				(ND-3.0)	
				Pulp: 2.77	
				(ND-10.0)	
Taiwan (lychee)	April–June	6	0.5	Peel: 6.8	Nakamura et
ruiwun (iyenee)	1987	Ū	0.5	(2–23.2)	al. 1989
				0 1 12 0	
				See as: 12.9	
				$\frac{(2.2-47.2)}{\text{Pulp} \cdot 0.9}$	
				1 uip. 0.7	
China (lychee)	April–June	1	0.5	Peel: 4.3	Nakamura et
	1987				al. 1989
				Seeds: 9.7	

Item sampled	Sampling period	Number of samples	Detection limit (µg/kg)	Mean concentration ¹ (µg/kg)	Reference	
United States (found in one sample of peanut butter and one sample of whiskey)	1988	231 samples (derived from US FDA market basket collection)	ns	Whiskey (80 proof): 2 Peanut butter: 11	Daft 1988	
United States					Rains and	
- flour	1980	22	5	807 (ND-4200)	Holder 1981	
- biscuits		22	0.5	36 (ND-260)	1101001 1901	
Japan (wheat; authors note that processing and market circulation would likely decrease levels)	1985	3	0.5	1.11 (0.74–1.70)	Konishi et al. 1986	
United States						
- flour, enriched		3		24		
- flour, unbleached pastry	1005	3	2	140	Clower et al.	
- meal, corn	1985	3	2	55	1960	
- wheat, whole grain red winter		3		167		
United States (cooked rice)	1984	4	0.4	2.5 (ND-8.3)	Clower et al. 1985	
Saskatoon, Saskatchewan (flour)	1984	10	ns	81 (4.1-405.3)	McKay 1986	

Abbreviations: ND, not detected; ns, not specified. ¹ Values in parentheses indicate range of concentrations when available. ² The limit of quantitation (LOQ) was obtained from a related total diet study of eight U.S. FDA market baskets during the period from April 1982 to April 1984 (Gunderson 1988b). The Gunderson (1988a) study incorporated the results of Gunderson (1988b) and included an additional two years of sampling (April 1984 to April 1986).

Appendix 6: Concentrations of 1,2-dibromoethane in soil

Location	Sampling period	No. of samples	Detection limit (ng/g) ¹	Mean Concentration (ng/g) ²	Reference
				3 m depth: 4.24×10^6 (dry weight)	
Site of a chemical plant, Ontario	1997	-	-	0.8 m depth: 1.19× 10 ⁴ (dry weight)	Environment Canada 2001b
				0.2–0.76 m depth: 80 (dry weight)	
Ontario regions (rural parkland, soil)	ca. 1993	59	MDL 4.0	0.032 ³ (0.012–0.390) ⁴ [dry weight]	OMEE 1993
Ontario (soil)	1986	5	MDL 0	$0.032^{3} (0.012-0.390)^{4}$ [dry weight]	OMEE 1993
Port Credit, Ontario (soil)	1987	8	MDL 0.2 [wet weight]	ND	Golder Associates 1987
Oakville/ Burlington, Ontario (soil)	1986	8	MDL 0.2–10 [wet weight]	ND	Golder Associates 1987

Abbreviations: MDL, method detection limit; ND, not detected.

¹ The MDL is defined as 3 times the within-run analytical standard deviation and is considered only an estimate that may vary with time (OMEE 1993).

² Values in parentheses indicate range of concentrations when available.

³ The concentration is the 97.5th percentile Ontario typical range value. This concentration is two standard deviations above the mean value.

⁴ The ranges are derived from the Ontario typical range model released in 1993 (to replace the previous "upper limit of normal" contaminant guidelines).

Appendix 7: Summary of health effects information for 1,2-dibromoethane

Endpoint	Lowest effect levels ¹ /Results
Laboratory anima	als and <i>in vitro</i>
Acute toxicity	Lowest oral LD ₅₀ (rabbit) = 55 mg/kg-bw (Rowe et al. 1952)
	Lowest inhalation $LC_{50}(rat) = 3080 \text{ mg/m}^3$ (Rowe et al. 1952)
	[Additional studies: Koptagel and Bulut 1998]
Short-term repeated-dose toxicity	Lowest oral LOEL (mice) = 125 mg/kg-bw per day based on increased cholesterol levels and increased <i>in vitro</i> phagocytosis of pooled cultured cells from 2–3 dosed animals at 125 mg/kg-bw per day and higher doses. Ethylene bromide (in corn oil) was injected intragastrically at doses of 100, 125, 160 or 200 mg/kg-bw per day for 14 days (n = 10 per treatment) (Ratajczak et al. 1994).
Subchronic toxicity	Lowest oral LOEL (mice) = 125 mg/kg-bw per day based on alterations in <i>in vivo</i> serum and hematology parameters and <i>in vitro</i> lymphocyte response. Ethylene bromide (in corn oil) was injected intragastrically at doses of 31.25, 62.5 or 125 mg/kg-bw per day, 5 days a week for 12 weeks (n = 6–9 per treatment) (Ratajczak et al. 1995). Lowest inhalation LOEC (rats) = 77 mg/m ³ based on epithelial hyperplasia of the nasal turbinates at 77 and 307 mg/m ³ . Rats were exposed to ethylene bromide at doses of 0, 3, 10 or 40 ppm (equivalent to 0, 23, 77 to 207 mg/m ³ as no MBCS 100(c).
	(n = 10 per treatment) (Nitschke et al. 1980).
Chronic toxicity/	Oral (gavage) carcinogenicity bioassay in rats: Males were exposed to a
carcinogenicity	time-weighted average of 0, 38 or 41 mg/kg-bw per day (5 days/week for up to 49 weeks). Females were exposed to 0, 37 or 39 mg/kg-bw per day (5 days/week for up to 61 weeks). Both sexes initially received 0, 40 or 80 mg/kg-bw per day of 1,2-dibromoethane, but, due to excessive mortality, the exposure levels and the overall duration of the study were reduced. In both sexes, there were significant increases in the incidence of squamous cell carcinomas of the forestomach in exposed groups (0/20 for both male and female controls, 45/50 for low-dose males, 33/50 for high-dose males, 40/50 for low-dose females, 29/50 for high-dose females). In males in the low-dose group, there was a significant increase in the incidence of hemangiosarcomas of the circulatory system (0/20 controls, 11/50 low dose); after time-adjusted analysis in high-dose females, there was a significant increase in the incidence of hepatocellular carcinomas (0/20 controls, 5/25 high dose) (NCI 1978).
	Oral (gavage) carcinogenicity bioassay in mice: Mice were exposed to time-weighted average doses of 0, 62 or 107 mg/kg-bw per day

Endpoint	Lowest effect levels ¹ /Results
	(5 days/week for 53 weeks). Mortality was high in all treated groups and due to this, all males and high-dose females were sacrificed at wk 78 (25 wks after dosing ceased). Low-dose females were sacrificed at wk 90. There were significant increases in the incidence of squamous cell carcinomas of the forestomach (males: vehicle control, 0/20; low dose, 45/50; high dose, 29/49; females: vehicle control, 0/20; low dose, 46/49; high dose, 28/50) and in alveolar/bronchiolar adenomas (males: control, 0/20; high dose, 10/47; females: control, 0/20; low dose, 11/43) (NCI 1978).
	[Additional study: Van Duuren et al. 1985 (drinking water): evidence of carcinogenicity was observed]
	Inhalation carcinogenicity bioassay in rats: Rats were exposed by inhalation to 0, 10 or 40 ppm (equivalent to 0, 77 or 308 mg/m ³) 6 h/day, 5 days/week, for 88–103 weeks). High mortality at the high concentration (90% in males, 84% in females) resulted in sacrifice of the remaining high-dose animals at wks 88 (males) or 91 (females). There were significant increases in the incidence of nasal cavity carcinomas at high doses (males: controls, 0/50; high dose, 21/50; females: controls, 0/50; high dose, 25/50) and adenocarcinomas at both doses (males: controls, 0/50; low dose, 20/50; high dose, 28/50; females: controls, 0/50; low dose, 20/50; high dose, 28/50; females: controls, 0/50; low dose, 20/50; high dose, 28/50; low doses (males: control, 0/50; low dose, 11/50; females: controls, 0/50; high dose, 5/50). There was a significant increase in the incidence of mammary gland fibroadenomas (controls, 4/50; low dose, 29/50; high dose, 24/50), and the highest-dose females exhibited significant levels of alveolar/bronchiolar adenomas combined with carcinomas (controls, 0/50; high dose, 5/47). Male rats had a significant increase in the incidence of tunica vaginalis mesotheliomas at both doses (controls, 0/50; low dose, 7/50; high dose, 25/50) and nasal cavity adenomatous polyps at the low dose (controls, 0/50; low dose, 18/50) (NTP 1982).
	Inhalation carcinogenicity bioassay in mice: Mice were exposed by inhalation to 0, 10, or 40 ppm (equivalent to 0, 77 or 308 mg/m ³) 6 h/day, 5 days/week, for 78–103 weeks). High mortality in both treated and control males resulted in sacrifice of all remaining males at wk 78. In females, high mortality was observed only at the high concentration (86%), and all remaining females at this concentration were sacrificed at wk 90. There were significantly increased incidences of alveolar/bronchiolar carcinomas (males: control, 0/41; high dose, 19/46; females: control, 1/49; high dose, 37/50) and adenomas (males: controls, 0/41; high dose, 11/46; females:
	controls, 3/49; high dose, 13/50) in the highest-dose groups of both sexes. In dosed females, there was also a significantly increased incidence of hemangiosarcomas of the circulatory system (controls, 0/50; low dose, 11/50; high dose, 23/50), subcutaneous fibrosarcomas (controls, 0/50; low dose, 5/50; high dose, 11/50), nasal cavity carcinomas (controls, 0/50; high

Endpoint	Lowest effect levels ¹ /Results
	dose, 6/50) and mammary gland adenocarcinomas (controls, 2/50; low dose, 14/50; high dose, 8/50) (NTP 1982).
	[Additional studies: Stinson et al. 1981; Wong et al. 1982: evidence of carcinogenicity was observed in both studies]
	Dermal carcinogenicity bioassay in mice: Female mice were given 0, 25 or 50 mg/mouse in acetone, dermally, 3 times a week for 440–594 days (equivalent to 357 or 714 mg/kg-bw per day, respectively; as per Health Canada 1994). There was a significant increase in the incidence of benign lung papillomas at both dose levels (low dose, 24/30; high dose, 26/30) and a significant increase in the incidence of combined squamous skin papillomas and carcinomas (3/30), as well as skin papillomas (5/30) at the high dose(Van Duuren et al. 1979).
	Lowest non-neoplastic oral (gavage) effect level (rats) = 38 (male) and 37 (female) mg/kg-bw per day, based on hyperkeratosis and acanthosis of the forestomach in females, degenerative changes in the liver, cortical cell degeneration of the adrenal gland and testicular atrophy in males (lowest dose tested, carcinogenic dose) (NCI 1978)
	Lowest non-neoplastic inhalation concentration $(rats) = 77 \text{ mg/m}^3$, based on toxic nephropathy and testicular degeneration in males, retinal atrophy and adrenal cortex degeneration in females and increases in hepatic necrosis in both sexes (lowest dose tested, carcinogenic dose; NTP 1982).
	[Additional studies: Stinson et al. 1981; NTP 1982; Wong et al. 1982]
Reproductive toxicity	Lowest oral (feed) LOEL (bulls) = 2 mg/kg-bw per day for 12 months (followed by 4 mg/kg-bw every 2 days for 10–12 months), based on reversible low sperm density, poor motility and altered spermatozoa morphology (Amir and Volcani 1965)
	Oral (gavage) at 38 mg/kg-bw per day for 49 weeks caused testicular atrophy in male rats (NCI 1978)
	[Additional study: Shivanandappa et al. 1987]
	Lowest inhalation LOEC (rats) = 77 mg/m ³ , based on testicular degeneration in males rats in a $88-103$ -week study (NTP 1982)
	Reproductive effects were reported in male or female rats following inhalation exposure to 0, 19, 39 or 89 ppm (equivalent to 146, 300 or 684 mg/m ³ as per IPCS 1996) in males or 0, 20, 39 or 80 ppm (equivalent to 154, 300, or 614 mg/m ³ as per IPCS 1996) in females for 10 or 3 weeks, respectively. In male rats, a reduction in testicular weight; decreased serum testosterone levels; atrophy of testes, epididymis, prostate and seminal vesicles; and changes in reproductive behaviour were reported only in the high-dose group. Also, female rats in the high-dose group showed abnormal

Endpoint	Lowest effect levels ¹ /Results
	estrous cycle until several days after cessation of exposure. Mortality
	occurred in both sexes in the high-dose group (Short et al. 1979).
Developmental	Lowest inhalation LOEC (rats) = 51.2 mg/m^3 , based on decreased
toxicity	maternal body weight and improved rotorod performance and T-maze
-	brightness discrimination acquisition in offspring (Smith and Goldman
	1983)
	[Additional study: Short et al. 1978]
Genotoxicity and	GENE MUTATION
related endpoints.	Positive results:
in vitro	Salmonella typhimurium TA98 (+/-S9) TA100 (+/-S9) TA100 (GSH-)
in viiro	(-S9 + GSH) TA100 (GSTA1-1 or GST1-1) (-S9) TA100W (Strr 8AGr)
	(-S9) TA102 (activation not specified) TA1530 (-S9) TA1535 +/-S9)
	TA1535 (GST1-1) ($-$ S9) TA2638 (activation not specified) G46 ($-$ S9)
	BA13 +/-S9) (Ames and Yanofsky 1971: Von Buselmaier et al. 1972: Brem
	et al. 1974: McCann et al. 1975: Rosenkranz 1977: Rannug and Beije 1979:
	Elliott and Ashby 1980: Shiau et al. 1980: Stolzenberg and Hine 1980: van
	Bladeren et al 1980 1981: Barber et al 1981: Principe et al 1981: Barber
	and Donish 1982: Kerklaan et al. 1983, 1985: Moriva et al. 1983: Buijs et
	al 1984: Dunkel et al 1985: Tennant et al 1986 1987: Hughes et al 1987.
	Zoetemelk et al. 1987: Ong et al. 1989: Roldán-Ariona et al. 1991: Zeiger et
	al 1992: Simula et al 1993: Novotná and Duverger-van Bogaert 1994.
	Thier et al 1996: Watanabe et al 1998)
	<i>Escherichia coli</i> WP2 (+/-S9) WP2/pKM101 (activation not specified)
	WP2 $\mu v r A/n K M 101$ (activation not specified) CHY832 (-S9) 343/286
	$(+/-S9)$ K12 $(+/-S9)$ K1201 $(-S9)$ K1211 $(-S9)$ $\mu\nu rB5$ (Scott et al. 1978)
	Hemminki et al 1980: Izutani et al 1980: Moriva et al 1983: Haves et al
	1984: Mohn et al 1984: Dunkel et al 1985: Foster et al 1988: Watanabe et
	al 1998)
	Bacillus subtilis TKJ5211, TKJ6321 (+S9) (Shiau et al. 1980)
	Streptomyces coelicolor (-S9, spot test) (Principe et al. 1981)
	Aspergillus nidulans (Scott et al. 1978: Principe et al. 1981)
	Neurospora crassa ad-3 (forward mutation) (De Serres and Malling 1983)
	Mouse L $5178Y$ (+/-S9) (Clive et al 1979: Tennant et al 1986–1987)
	Chinese hamster CHO-K1($+/-$ S9) (Tan and Hsie 1981: Brimer et al. 1982)
	Human cell line AHH-1 TK6 (-S9) (Crespi et al. 1985)
	Human cell line EUE (–S9) (Ferreri et al. 1983)
	<i>E. coli</i> lacZ reversion assay (Josephy et al. 2006)
	Negative results:
	Salmonella typhimurium TA98 (+/-S9) TA100 (+/-S9) TA1537 (+/-S9)
	TA1538 (\pm /-S9) E503 (Brem et al. 1974: Alper and Ames 1975: Shiau et
	al 1980: Principe et al 1981: Wildeman and Nazar 1982: Moriva et al
	1983: Dunkel et al. 1985: Tennant et al. 1986)
	Serratia marcescens a ²¹ (-S9) (Von Buselmaier et al. 1972)
	Escherichia coli $343/113$ (-S9) (Mohn et al. 1984)
	Strentomyces coelicolor (-S9 nlate method) (Princine et al. 1981)
	Sirepromyces coefficion (159, plate method) (1 metpe et al. 1981)
	UNSCHEDULED DNA SYNTHESIS

Endpoint	Lowest effect levels ¹ /Results
	Positive results:
	Rat hepatocytes (Williams et al. 1982; Tennant et al. 1986; Working et al. 1986)
	Rat spermatocytes (Working et al. 1986)
	Opossum lymphocytes (Meneghini 1974)
	Human lymphocytes (+/-S9) (Perocco and Prodi 1981)
	Mouse (C3H1×101)F1 germ cens (Sega and Solomayor 1980)
	SISTER CHROMATID EXCHANGE
	Chinese hamster V79 cl-15 (-S9) (Tezuka et al. 1980)
	Chinese hamster ovary (+/-S9) (Tennant et al. 1980)
	Human lymphocytes (-S9) (Tucker et al. 1984; Ong et al. 1989)
	CHROMOSOMAL ABERRATIONS
	Positive results: Chinaga hamatar V70 al 15 (-S0) (Tazuka at al 1080)
	Chinese hamster ovary (+/-S9) (Tennant et al. 1980) Chinese hamster ovary (+/-S9) (Tennant et al. 1987; Ivett et al. 1989)
	MICRONUCLEI INDUCTION
	Human lymphocytes (Channarayanna et al. 1992)
	Tuman Tymphocytes (Chaimarayappa et al. 1992)
	DNA DAMAGE
	Positive results:
	<i>Escherichia coli</i> polA1–/polA+(-S9) (Brem et al. 19/4) Human pasal mucosa cells, rat ethnoidal mucosa, rat pasal mucosa cells
	(Holzer et al. 2008)
	Negative results:
	Bacilis subtilis TKJ5211, TKJ6321 (+/-S9) (Shiau et al. 1980)
	SOS INDUCTION
	Positive results:
	Salmonella typhimurium TA1535/pSK1002 (+/-S9), NM5004 expressing
	GS1 5-5 (Ong et al. 1987; Oda et al. 1996) Escherichia coli (Obta et al. 1984: Quillardet et al. 1985)
	Escherichia con (Onta et al. 1964, Quinaidet et al. 1965)
	Negative results:
	Salmonella typhimurium TA1535/pSK1002 (-S9) (Oda et al. 1996)
	MITOTIC GENE CONVERSION
	Positive results:
	Saccharomyces cerevisiae ade2, trp5 (Fahrig 1974)
	SOMATIC SEGREGATION
	Positive results:
	Aspergillus nidulans diploid 35×17 (-S9) (Crebelli et al. 1984)

Endpoint	Lowest effect levels ¹ /Results
	CELL PROLIFERATION
	Positive results:
	Human lymphocytes (Channarayappa et al. 1992)
	DNA STRAND BREAKS
	Positive results:
	Rat hepatocytes (Sina et al. 1983)
	Rat testicular cells (Bradley and Dysart 1985)
	Rat and human testicular cells (Bjørge et al. 1996)
	DNA BINDING
	Cali thymus DNA (Artellini et al. 1984; Colacci et al. 1985; Prodi et al.
	$\frac{1980}{1}$
	Kat hepatocytes (Inskeep et al. 1986; Cmarik et al. 1990)
	Human nepatocytes (Cmarik et al. 1990)
	Nogative regults:
	Regative results. Escherichia coli $O(12) (\pm 1/50)$ and mouse Ehrlich assistes $(\pm 1/50)$ (Kubinski
	et al. 1981)
	et al. 1901)
	CELL TRANSFORMATION
	Positive results:
	Balb/c 3T3 mouse cells (Perocco et al. 1991: Colacci et al. 1995)
	Negative results:
	Balb/c 3T3 mouse cells (-S9) (Tennant et al. 1986)
Genotoxicity and	GENE MUTATION
related endpoints:	Positive results:
in vivo	Drosophila melanogaster (Graf et al. 1984; Ballering et al. 1993)
	Salmonella typhimurium G46 host-mediated (Von Buselmaier et al. 1972)
	Negative results:
	Serratia marcescens host-mediated (Von Buselmaier et al. 1972)
	Silk worm (Sugiyama 1980)
	RECOMBINATION
	Positive results: $D = \frac{1}{2} \frac{1}{$
	Drosophila melanogaster (Graf et al. 1984; Ballering et al. 1993)
	SEV I INVED DECESSIVE LETHAL MUTATIONS
	Desitive results:
	Drosophila melanogaster (Vogel and Chandler 1974. Kale and Raum
	1979a 1979b 1981 1982 1983 Yoshida and Inagaki 1986 Ballering et al
	1993, 1994; Foureman et al. 1994; Kale and Kale 1995)
	CHROMOSOMAL ABERRATIONS
	Negative results:
	Mouse (intraperitoneal) bone marrow (Krishna et al. 1985) (IARC reports
	weakly positive) (IARC 1999)
Endpoint	Lowest effect levels ¹ /Results
----------	---
-	Mouse (intraperitoneal) bone marrow (NTP 1993)
	DNA STRAND BREAKS
	Positive results:
	Rat hepatocytes (Nachtomi and Sarma 1977; Kitchin and Brown 1994)
	Mouse hepatocytes (White 1982; Storer and Conolly 1983)
	Rat testicular cells (Bradley and Dysart 1985)
	MICRONUCLEI
	Positive results:
	Mouse (peripheral blood) (Witt et al. 2000)
	Negative results:
	Mouse (Krishna et al. 1985; Asita et al. 1992)
	DNA BINDING
	Positive results:
	Mouse (liver, stomach, kidney, lung) (Arfellini et al. 1984; Prodi et al.
	1986)
	Mouse hepatocyte DNA (Kim and Guengerich 1990)
	Mouse (liver, kidney) (Watanabe et al. 2007)
	Rat (liver, stomach, kidney, lung) (Arfellini et al. 1984; Prodi et al. 1986)
	Rat hepatocyte DNA (Inskeep et al. 1986; Kim and Guengerich 1990)
	Rat (liver, kidney) (Watanabe et al. 2007)
	SPECIFIC LOCUS TEST
	Negative results:
	Mouse (Russell 1986; Barnett et al. 1992)
	SISTER CHROMATID EXCHANGE
	Negative results:
	Mouse (intraperitoneal) bone marrow (Krishna et al. 1985)
	Mouse (intraperitoneal) bone marrow (NTP 1992)
	DOMINANT LETHAL
	Negative results:
	Rat (Short et al. 1979; Teramoto et al. 1980; Teaf et al. 1990)
	Mouse (Epstein et al. 1972; Teramoto et al. 1980; Barnett et al. 1992)
	DNA REPAIR EXCLUSIVE OF UNSCHEDULED DNA SYNTHESIS
	Negative results:
	Mouse hepatocytes (White et al. 1981)
	UNSCHEDULED DNA SYNTHESIS
	Positive results:
	Rat hepatocytes (Working et al. 1986)
	Negative results:
	Rat spermatocytes (Working et al. 1986; Bentley and Working 1988)

Endpoint	Lowest effect levels ¹ /Results
	DNA DAMAGE
	Positive results: Mouse (stemach liver kidney bladder lung) (Secoli et al. 1008)
Humans	Nouse (stollach, liver, kidney, bladder, lung) (Sasaki et al. 1998)
Acute toxicity	Estimated fatal dose in adult male and female (human) = 1.5 ml or 3240 mg (46 mg/kg-bw for a 70-kg person). Effects observed included nausea, vomiting, abdominal pain and signs of hepatotoxicity, nephrotoxicity, nervous system toxicity and cardiotoxicity in male and female patients (Singh et al 2007 - review of 64 cases of acute 1,2-dibromoethane poisoning).
	Estimated inhalation lethal concentration (human) = 154 mg/m^3 for more than 30 min (IPCS 1996)
	[Additional studies: Alexeeff et al. 1990 ; Peoples et al. 1978; Letz et al. 1984; Jacobs 1985; Sarawat et al. 1986; Singh et al. 1993; Prakash et al. 1999; Raman and Sain 1999; Mehrotra et al. 2001]
Chronic toxicity/ carcinogenicity	Mortality assessed in employees occupationally exposed to 1,2-dibromoethane in two production units while working as still and reactor operators (level of exposure was not provided in secondary accounts). In the first production unit, there were 2 deaths from malignant neoplasms (3.6 expected), and in the second production unit, there were 5 deaths from malignant neoplasms (2.2 expected). However, employees of the second production unit were also exposed to other chemicals, and overall there was no increase in total deaths or malignant neoplasms with increased exposure (Ott et al. 1980).
	[Additional study: Ter Haar 1980]
Reproductive and developmental toxicity	Lowest inhalation LOEC = 0.46 mg/m ³ based on significantly decreased sperm velocity and semen volume in male forestry workers (occupational time-weighted average) in male forestry workers. Forestry workers engaged in applying or spraying of 1,2-dibromoethane emulsion (4% 1,2-dibromoethane by volume) were examined following short-term inhalation and dermal exposure (Schrader et al. 1988; IPCS 1996).
	Male forestry workers conducting fumigation (n = 46) with 1,2-dibromoethane for 5 years, showed significant decreases in sperm count, number of viable sperms and increase in sperms with abnormal morphology. 1,2-Dibromoethane concentration ranged from a geometric mean of 88 ppb to peak concentration of up to 262 ppb (equivalent to 0.68 mg/m^3 to 2.0 mg/m^3 as per IPCS 1996) for 8-hr time-weighted average. The authors did not report exposure to any other chemicals in the forestry workers engaged in the application or spraying activities (Ratcliffe et al. 1987).
Constavisity or d	[Additional studies: Ter Haar 1980; Wong et al. 1985; Dobbins 1987;Schrader et al. 1987]
Genotoxicity and	negative results:

Endpoint	Lowest effect levels ¹ /Results
related endpoints	Chromosomal aberrations and sister chromatid exchange were not detected in men who worked in papaya-packing plants and used 1,2-dibromoethane to fumigate the fruit. These workers were exposed to mean concentrations ranging from 0.12 to 1.35 mg/m ³ (Steenland et al. 1986).
	[Additional study: Steenland et al. 1985]

 1 LC₅₀ = median lethal concentration; LD₅₀ = median lethal dose; LOEC = lowest-observed-effect concentration; LOEL = lowest-observed-effect level.